

# COMPARATIVE STUDIES OF THE PERFORMANCE OF FILTER MEDIA MADE USING BIOCHAR AND ACTIVATED CARBON IN GREYWATER REMEDIATION

**BY**

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# AHMADU BELLO UNIVERSITY, ZARIA, NIGERIA

**MAY, 2017**

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**BY**

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**B.ENG AGRIC & ENV. RES (UNIMAID, 2010) M.Sc/ENG/41914/12-13**

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# DEPARTMENT OF WATER RESOURCES AND ENVIRONMENTAL ENGINEERING,

**AHMADU BELLO UNIVERSITY, ZARIA, NIGERIA**

# MAY, 2017

# DECLARATION

I declare that the work in this dissertation titled ―COMPARATIVE STUDIES OF THE PERFORMANCE OF FILTER MEDIA MADE USING BIOCHAR AND ACTIVATED

CARBON IN GREYWATER REMEDIATION‖ is the result of my own investigation.

The information derived from the literature has been duly acknowledged in the text and a list of references provided.

SalihuWamdeo SAMUEL

(M.Sc/ENG/41914/2012-2013) Signature Date

# CERTIFICATION

This dissertation titled ―COMPARATIVE STUDIES OF THE PERFORMANCE OF FILTER MEDIA MADE USING BIOCHAR AND ACTIVATED CARBON IN

GREYWATER REMEDIATION‖ By SalihuWamdeo SAMUELmeets the regulations governing the award of the degree of Master of Sciences (M.Sc) of Ahmadu Bello University, Zaria, and approved for its contribution to knowledge and literary presentation.

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# DEDICATION

This dissertation is dedicated to the loving memory of my late father Mr. Samuel S. Wamdeo.

## ACKNOWLEDGEMENT

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## ABSTRACT

The problem of water Scarcity, water pollution and cost of accessing water for domestic needs have become growing concerns to the ever increasing global population. Waterwaterremediation has long been accepted as a viable option to mitigate water scarcity. This research examines the potentials of an environmentally friendly technique of using local but modified tropical plant materials in remediation of greywater for re-use. Column adsorption experiment was conducted using synthesized and characterized Biochar and Activated Carbon made from Shea tree (*Vitellariaparadoxa*), African Mahogany tree (*KhayaSenegalenses*) and the Fig tree (*FicusSycomous*) as filter media in constructed filter columns made from polyvinyl chloride pipes. Greywater Samples from Ahmadu Bello University Zaria‘s student hostel were passed through these filters at a hydraulic loading rate of 0.033m3/m2/day for a period of 21 days. The parameters assessed in this experiment were Electrical Conductivity (EC), Biological and Chemical Oxygen Demand (BOD and COD), Ammonium Nitrogen (NH4-N), Nitrate Nitrogen (NO3-N), Total Nitrogen (Tot-N), available Phosphate (PO4-P), Total Phosphorus (Tot-P) and pH. The results obtained showed high treatment efficiencies for both Activated Carbon and Biochar on all tested parameters except for NH4-N in which AC had a low efficiency of 49% at one instance. Activated Carbon had (85 – 95%) in remediating organic matter, (61 -81%) in remediating EC, (39 – 89%) in remediating NH4-N, (93 – 100%) for NO3-N, 92 – 99% for Total Phosphorus. Biochar showed 88 – 92% efficiency in remediating Total Phosphorus, 91 – 99% for NO3-N, 68 – 89% for NH4-N, 85 – 90% for EC and 76 – 90% for organic matter removal. The factors of time, particle size, activation status and parent material used in the filters were used to determine the efficiency of the filters and the results showed that time, parent material used, and activated status of the filters had significant effect on the performance of the filters while differences in particle sizes had little effect on the performance. It could be concluded therefore that biochar compared favourably with activated carbon in the remediation of greywater, which is more complex to synthesize and cost more to produce. In addition, because of the ease of production and availability of raw materials in the tropics, biochar can be used to remediate greywater and replace Activated Carbon in some water treatment systems.

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LIST OF ABBREVIATIONS AND ACRONYMS

ABU - Ahmadu Bello University

AC - Activated Carbon ANOVA - Analysis of Variance

BOD - Biological Oxygen Demand CEC - Cation Exchange Capacity COD - Chemical Oxygen Demand DOS - Department of Statistics

EC - Electrical Conductivity

MCM - Million Cubic Meters

NA - Not Available

NH4-N - Ammonium Nitrogen NS - Not Significant

NO3-N - Nitrate Nitrogen

PO4-P - Phosphate Phosphorus

PPRC - Pollution Prevention Resource Centre PS - Particle Size

PVC - Poly Vinyl Chloride

SOC - Synthetic Organic Compounds TOT-N - Total Nitrogen

UN - United Nations

VOC - Volatile Organic Compounds

WREE - Water Resources and Environmental Engineering WHO - World Health Organisation

## CHAPTER ONE INTRODUCTION

* 1. **Background of the study**

With the continuous annual increase in the world population (lowest increase being 1.1%) there is need to manage our natural resources. Water, which is one of these basic resources for human survival and socioeconomic development, needs to be managed efficiently. The United Nations has estimated that the use of water has increased six fold in the 20th century and that by the year 2025, about 1.8 billion people will have absolute water scarcity conditions (with an annual water supply of less than 500m3 per capita) and also that two third (2/3) of the world population will live under water stress (with an annual water supply of less than 1700m3 per capita) (U.N-Water Report, 2006). This population increase will have adverse effects on developing countries where 82% of the world live and experience water scarcity (Dalahmeh, 2013). There is therefore need to properly manage and utilize existing water resources especially in the developing countries.

The reality of water scarcity, coupled with the increased potential health risk posed by greywater pollution necessitates a swift solution hence the idea of using locally available materials like biochar as a means of improving water quality, sanitation and reducing environmental problems has become attractive.This study seeks to find simple inexpensive and locally available materials like charcoal and sand to solve environmental problems posed by water scarcity and water pollution.

Wood from locally available tropical trees waspyrolyzed at a temperature of 650oC and the biochar derived wascharacterized and treated. Its ability to adsorb certain major pollutants from the collected greywater was determined.

Greywater or sullage is defined as wastewater generated from kitchens and wash-hand basins, showers and baths, which, because it is nearly as clean as potable water, can be recycled onsite for uses such as toilet flushing, landscape irrigation and constructed wetlands (Barker and English*,* 2011).Biochar refers to biomass-derived char product most often used as cooking fuel (Kearns*et al,* 2014). It is a name for charcoal when it is used for a particular purpose such as soil amendment, water treatment, etc. (Kearns, 2012).Pyrolysis is a thermochemical decomposition of organic materials at elevated temperature in the absence of oxygen. It involves the simultaneous change in both chemical and physical compositions of the organic materials. The process is irreversible. Pyrolysis produces gas, liquids and a solid product rich in carbon.

## Statement of the research Problem

Greywater, which accounts for approximately 50-80% of total household wastewater mostly ends up in receiving water bodies (Li *et al.,* 2009). The provision of inadequate sanitation and wastewater disposal facilities leads to environmental and public health problems which is responsible for 1.8 million deaths annually (Corcoran *et al*., 2010; WHO Report, 2009). Although centralized wastewater systems are common in industrialized countries, they are complex and expensive to construct and operate. This however is not a viable option for most communities in developing countries. There is therefore the need to source for simple and affordable environment friendly techniques to treat greywater.

## Justification of the Study

Previous studies and designs utilize activated carbon as adsorbents in water treatment systems. Recent studies however showed that biochar especially those with high lignin

content when pyrolyzed at temperatures above 450oC show similar molecular structure like that of activated carbon and can therefore be a possible replacement to Activated Carbon (McLaughlin *et al.*, 2009; Kearns, 2012). This study aims to experimentally determine the effectiveness of biochar in greywater remediation. Biochar is cheaper, available and more environmentally friendly when compared to activated carbon, ion exchange resins, membrane filtration, reverse osmosis and nano filtration (Kearns, 2012). Biochar production techniques are also cheap and easy for local communities and so, this might be a better treatment option for community wastewater remediation especially in developing countries.

## Aim and Objectives

This research aims to determine the effectiveness and efficiency of biochar derived from different lignin containing trees; *Vitellariaparadoxa* (Shea Tree), *Khayasenegalenses* (Mahogany), *Ficussycomous*(Fig) in remediating greywater.

The research has these following specific objectives.

* + 1. To produce biochar and activated carbon from different parent material
    2. To use the biochar and activated carbon to make filters with physical properties like those of sand filters.
    3. To determine the efficiency of the prepared filters in remediating chemical and physical parameters; pH, Electronic Conductivity (EC), Ammonium Nitrogen (NH4-N), Total Nitrogen (Tot-N), Phosphate Phosphorus (PO4 -P), Nitrate Nitrogen (NO3-N), Biological Oxygen Demand and Chemical Oxygen Demand (COD) from greywater.
    4. To compare the performance of the biochar with activated carbon made from same parent materials in greywater remediation.
    5. To determine the effect of particle size, time and parent material in adsorbing the stated parameters.

## Scope and Limitation of the study

This studywas to determine the effectiveness of biochar from three specifically identified tropical lignin containing trees (*Vitellariaparadoxa*, *Khayasenegalenses*, *Ficussycomous)* in remediating greywater fromSuleiman Hostel of Ahmadu Bello University Zaria (A.B.U). Physical and chemical parameters tested were: pH, Electrical Conductivity (EC), Amonium Nitrogen (NH4-N), Total Nitrogen (Tot-N), Phosphate- Phosphorus(PO4-P), Total Nitrogen, Nitrate Nitrogen (NO3-N), Chemical Oxygen Demand (COD) and(BOD)Biological Oxygen Demand. The constant mass of Carbon used throughout the experiment and the variation in daily concentration of pollutants in the greywater source however served as limitations in this study. It was addresses by adding an extra column which served as control to monitor changes in the greywater. This research was limited to Greywater and not industrial or other domestic wasterwater.

CHAPTER TWO

## LITERATURE REVIEW

* 1. **Greywater**

Morel and Diener, (2006) defined greywater as household wastewater that does not come from a toilet, meaning it is wastewater that does not contain urine or feces. Ludwig,(2012) and Mungai, (2008) reported that greywater makes up 50% to 80% of a household's wastewater. They further stressed that the quantity of greywater produced by a household depends above all on practices of the household in question which are directly influenced by the amount of freshwater available, the cost and supply route of that freshwater, the number of people living in the household, their ages and gender and the living conditions in the nearby area.

Winblad and Simpson*,* (2004) observed that the quality of greywater, e.g. level of contamination, varies depending on the household‗s water habits and the chemicals used. Murphy, (2006) noted that freshwater shortages and conservation will not only reduce the quantity of greywater produced, but also its quality, resulting in greywater that is more concentrated due to less dilution . Just as greywater quality and quantity varies from household to household, it likewise varies within a household depending on the source of the specific greywater. Morel and Diener, (2006) classifiedgreywater into three different types: kitchen, bathroom and laundry.

Oron,*et al.,*(2014) reported that domestic water use industrialized countries approximates amounts to 100-150 l/c/d (litre/*capita*/day), of which 60-70% is transformed into greywater, while most of the rest is consumed for toilet flushing and released as blackwater. Oron,*et al.,*(2014) iterated thatgreywater reuse for toilet flushing can reduce the in-house net water consumption by 40-60 l/c/d, leading to 10-20% reduction of the urban water consumption, which is significant especially under water

scarcity situation.Oron, *et al.,* (2014) further advised that additional reduction can be achieved by reusing greywater for garden irrigation which is a considerable water consumer in some semi arid regions (Australia, California, Israel, etc.).Friedler and Galil,(2003) showed that by the year 2023, with moderate penetration ratio of greywater reuse systems of 20-30% (percentage of houses having greywater reuse units installed), reuse of greywater in the urban sector in Israel could save 30-55 millioncubic meters per year, 25-45 and 5-10 Million Cubic Meters per year in toilet flushing and garden irrigation respectively. With a projected population ten million people, this amounts to about 5% of the total future urban water demand in the country, which they observed equals the capacity of a medium size seawater desalination plant.

## Greywater from Bathroom

Bathroom greywater is regarded as the least contaminated greywater source within ahousehold. It contains soaps, shampoos, toothpaste, and other body care products.Bathroom greywater also contains shaving waste, skin, hair, body-fats, lint, and traces ofurine and faeces. Greywater originating from shower and bath may thus be contaminatedwith pathogenic microorganisms(Morel and Diener, 2006).

## Greywater from Laundry

Laundry greywater contains high concentrations of chemicals from soap powders (suchas sodium, phosphorous, surfactants, Nitrogen) as well as bleaches, suspended solids and possibly oils, paints, solvents and non-biodegradable fibres from clothing. Laundry greywater can contain high amounts of pathogens when nappies are washed(Morel and Diener, 2006).

## Greywater from Kitchen

Kitchen greywater contains food residues, high amounts of oil and fat, including dishwashingdetergents. In addition, it occasionally contains drain cleaners and bleach. Kitchen greywateris high in nutrients and suspended solids. Greywater from dishwashers may be very alkaline (dueto builders), it also shows high suspendedsolids and salt concentrations (Morel and Diener,2006).

## Greywater Remediation

Morel and Diener, (2006) suggested that greywater should be regarded as a valuable resource and not a waste as it has great potential to reduce water stress currently faced in regions around the world. They further iterated that greywater reuse may lead to considerable economic benefit.

The United State Department of Statistics (US-DOS), (2001) also reported that in Amman, Jordan an extensive survey of urban farmers revealed that percent use greywater to irrigate gardens. Faruquiand Al-Jayyousi,(2002) similarly iterated that household treating and re-use of greywater locally may bring an annual benefit of USD 376 accounted for by increasing product yield as well as reduced water and fertilizer cost.

With a typical production of 90-120l/p/d (litres per person per day), Morel and Diener, (2006) suggested the need to remediate greywater.Igboro*et al.,* (2015) outlined the benefits of reusing greywater which include domestic and gardening purposes.

Allen *et al.,* (2010) suggested different methods of greywater remediation amongst which are: disinfection using chlorine, ozone or ultraviolet light, use of anaerobic

biological treatment, use of membrane bio-reactor, use of sand filtration and the use of activated carbon filters.

Igboro*et al.,* (2016) stated that some basic pollutants of greywater: BOD, Turbidity, Total Solids and E. coli can be improved using locust bean seed, leaves and bark.

## Biochar

Pollution Prevention Resource Centre (PPRC),(2014)defined biochar as ‗a name for charcoal when it is used for a particular purpose such as soil amendment, water treatment, carbon sequestration, bio-bags and energy production‘.Harris, (1999) quoted that the biochar production process mirrors that of charcoal which can be considered as one of the oldest invented industrial technologies of Homo sapiens. However, it can be differentiated from charcoal and other similar materials by considering the fact that biochar is synthesized completely with the intention of being applied to the soil for improving the soil fertility, carbon sequestration and other uses (Harris, 1999).

Pollution Prevention Resource Centre (PPRC), (2014) established that biochar‘s incredible porosity and surface area gives it a high capacity to adsorb a wide variety of contaminants from water. They further suggested that biochar can be thought as a cheaper version of activated Carbon which can be used in many of the same applications. They presented a research by PPRC in the Pacific North West showed that biochar can effectively reduce contaminants including heavy metals, Volatile Organic Compounds (VOCs), Synthetic Organic Compounds (SOCs), Oxygen Demand, Phosphorus, Phosphates, Ammonim, Nitrates, Nitrogen, Total Suspended Solids (TSS) and polyaromatic hydrocarbons.

## Biochar Production

The process of producing biochar is known as pyrolysis. This refers to the thermal decomposition of biomass under low or restricted oxygen conditions. The pyrolyticprocess parameters such as temperature, heating rate, pressure, energy values of bio-oils and physico-chemical properties all have effect on the production of biochar (Clay and Malo, 2012).

Biochar yield as well as, its chemical and physical characteristics of biochar depends on thenature of the feedstocks used (woody against herbaceous) and operating conditions and environmentof the pyrolysis unit (low against high temperature, residence time; slow vs. fast pyrolysis, heatingrate and feedstock preparation, etc.). Baldock and Smernik, (2002) reported that the wide range of process parameters leads to theformation of biochar products that vary considerably in their elemental and ash composition,density, pore size, surface chemical properties, water and ion adsorption and release, surfacearea, pH and uniformity of biochars‘ physical structure. Similar studies were also reported in literature such as Antal andGronli, (2003);Downie*et al.,* (2009);Amonette and Joseph,(2009);Krull*et al.*,(2009); Chan and Xu, (2009).

The biochar yield from the pyrolysis of biomass is influenced by the pyrolysistemperature, its lignin, cellulose, and hemicellulose content and to a lesser extent the extractiveconcentrations of the feedstock according to McKay and Roberts, (1982), Antal and Gronli, (2003). Demirbas, (2004) reported that woodybiomass with high lignin contents typically produce greater char yields than those derived fromherbaceous feedstocks. A common feature among the various pyrolysisprocesses is that C content shows a consistent increase with increasing temperature (Antal andGronli, 2003; Schnitzer*et al*., 2007; Zabaniotou*et al*., 2008; Lehman and Joseph, 2009). Biochars with largeamounts of poly-condensed aromatic structures are obtained by pyrolyzingfeedstocks attemperatures between 400 and 600oC (Antal and Gronli,

2003; Amonette and Joseph, 2009).Temperatures above 500oC commonly produce chars with Carbon contents greater than 80%, attemperatures between 400 and 500oC biochars have C contents that range from 60 to 80%, wherebiochars produced at low temperatures (< 350oC) have C content that vary from 15 to 60%(Lehman and Joseph, 2009). For a detailed description of the thermal degradation process (dehydration,pyrolysis, graphene formation and carbonization) that transforms biomass.

## Biochar for Water Treatment

Liang *et al.,* (2006) reported that biochar has the potential to increase Cation Exchange Capacity (CEC) in soils and he attributed this characteristics to the structural nature of biochar. Steinbiss*et al.,*(2009) reported that biochar may enhance microbial growth by providing pores as refuge to micro-organisms or by providing larger surface area for take-up.

Khan*et al.,* (2010) published that organic materials generally possess some adsorption capacity and their high carbon content provides resistance to decomposition. They also stated that charring these organic material usually increases their adsorption capacity by providing larger surface area for adsorption.Agoyi, (2015) studied the adsorption performance of cow bone char in a fix bed system and reported that cow bone adsorbent was useful in treating Lead and Chromium from contaminated wastewater.

Taghizadeh-Toosi*et al.,*(2012) published that biochar had been found to adsorb gaseous ammonia and make it readily available to plants.

Berger, (2012) pointed out that biochar which is an organic material pyrolysed/charred often by means of low cost techniques might be an interesting alternative to replace the industrial activated Carbon for greywater cleaning. She further evaluated and compared biochar with activated Carbon in cleaning synthesized greywater and published that

biochar performed more effectively in removing Total Phosphorus and Phosphates whereas both media had average performance in cleaning organic matter primarily Chemical Oxygen Demand (COD) and Methylene Blue Active Substances(MBAS) from the synthesized greywater. Similarly, Dalahmeh, (2013) also studied the performance of pine made biochar, activated Carbon and sand filters in removing nutrients from synthetic greywater. She published that the performance of the filters were more influenced by the Hydraulic and Organic loading rates of the previous runs in the experiment.

Bernd *et al.,* (2013) also reported that biochar has the possibility of replacing coal based activated Carbon as adsorbent for contaminants and pathogens in water purification systems. They further hypothesized that biochar could be used for pathogen removal from wastewater while at the same time being loaded with nutrient and contaminants.

Ghezzehei*et al.,* (2014) published that biochar can adsorb up to 20 – 43% of ammonium and 19 – 65% of phosphates in flushed dairy manure in 2 hours. They also suggested the potential of biochar in recovering nutrient from dairy waste water and improving soil fertility if the enriched biochar is returned to the soil.Foereid, (2015) noted that biochar has so far been mainly promoted for soil amendment and also a good adsorbent. They suggested that biochar may also hold promise in low cost wastewater treatment than activated Carbon which has been used for a long while. They further suggested that biochar could potentially be used to adsorb nutrient from wastewater and then the loaded biochar or mixture of sludge and biochar could be added to soil.

Huggins *et al.,* (2016) performed comparative batch sorption and column studies between Granular Activated Carbon (GAC) and biochar for wastewater treatment. They reported that biochar had greater adsorption capacity at high initial concentration of

Total COD, Phosphates and Ammonia. They also reported similar performance of biochar for bed column studies. They attributed the performance of biochar to its macro porous structure as compared to the microporous structure of Granular Activated Carbon.

## Introduction to Parent Material Used for the Biochar production

* + 1. **Khaya*senegalensis***

Khaya*senegalensis* is a species of tree in the Meliaceae family that is native to Africa. Common names include African mahogany, dry zone mahogany, Gambia mahogany, khaya wood, Senegal mahogany, cailcedrat, acajou, djalla, and popularly known as Mahogany in Northern Nigeria.

African mahogany is a medium-sized tree which can grow up to 15–30 m in height and 1 m in diameter. The bark is dark grey to grey-brown while the heartwood is brown with a pink-red pigment made up of coarse interlocking grains. The tree is characterized by leaves arranged in a spiral formation clustered at the end of branches.

* + 1. **Vitellaria*paradoxa***

Vitellaria*paradoxa*commonly known as shea tree, shi tree or simply vitellaria, is a tree of the Sapotaceae family. It is the only species in genus Vitellaria, and is indigenous to Africa. Vitellaria*paradoxa* is a small to medium-sized tree (min. 7) 10-15 (max. 25) m high; much branched, dense, spreading, round to hemispherical crown. In mature trees, the bole is short, usually 3-4 m but exceptionally 8m, with a diameter ranging from 0.3 to 1 m, but most frequently 0.6 m. Bark conspicuously thick, corky, horizontally and longitudinally deeply fissured; protects older trees against bush fires. Slash pale pink, secreting white latex, as do broken twigs or petioles. Leaves in dense clusters, spirally

arranged at the end of stout twigs. They are covered by thick bark showing numerous leaf scars. Petioles 5-15 cm long, leaves oblong. Juvenile leaves rust-red and pubescent, later coriaceous, glabrous and dark green, shining, 12-25 cm long and 4-7 cm wide, leaf margin wavy and bent (Orwa*et al.*, 2009).

* + 1. **Ficus*sycomorus***

Ficussycomorus locally known as Fig or Cedia (Hausa) is a fig species that has been cultivated since ancient times. Ficussycomorus grows to 20 m tall and has a considerable spread with a dense round crown of spreading branches. The leaves are heart-shaped with a round apex, 14 cm long by 10 cm wide, and arranged spirally around the twig. They are dark green above and lighter with prominent yellow veins below, and both surfaces are rough to the touch. The petiole is 0.5–3 cm long and pubescent. The fruit is a large edible fig, 2–3 cm in diameter, ripening from buff-green to yellow or red. They are borne in thick clusters on long branchlets or the leaf axil. Flowering and fruiting occurs year-round, peaking from July to December. The bark is green-yellow to orange and exfoliates in papery strips to reveal the yellow inner bark. Like all other figs, it contains a latex (Zohary and Maria, 2000).

## Lignin

Lignin is a heterogeneous and highly cross-linked macromolecule that represents the second most abundant natural polymeric material on earth (Guerra *et al.*, 2006). Lignin is that which makes plants ―woody‖ as the name comes from the neuter Latin noun

―lignum‖ meaning ―wood‖ (Harkin, 1969). Kearns, (2012) suggested that the adsorption of cellulose material is a primary function of its lignin content. Lignin is a complex macromolecule, and hence, it has been stated many times that there is no single method for the accurate determination of the quality or quantity of lignin (Davin

*et al.*, 2008;Brinkmann*et al.,* 2002).Aygun*et al.,* (2003) noted that the mechanical strength is a characteristic used for defining the quality of biochar and activated carbon as it relates to its ability to endure wear and tear during use. For this reason however, agricultural wastes, such as nut shells and fruit stones are of interest as activated carbons because of their high mechanical strength and hardness. These properties can be explained by high lignin and low ash contents.

## Adsorption

The word adsorption was first coined by a German Physicist Heinrich Kayser in 1881. It is the adhesion of atoms, ions or molecules from a gas, liuid or dissolved solid to a surfact. The process creates a film of the adsorbate on the surface of the adsorbent. Dabrauski, (2003) referred to adsorption as the change in concentration of a given substance at the interface as compared with the neighboring phases. Depending on the type of phases in contact, adsorption can be classified as:

* Liquid – gas
* Liquid – liquid
* Solid – Liquid
* Solid – Gas.

## The Science of Adsorption

Taking the gas solid phase as an example, the basic concept in adsorption occurring at every interface is the real adsorption system which is defined as an equilibrium one including the adsorbent being in contact with the bulk phase and the interfacial layer. This layer consist of two region, the part of gas residing in the force field of the solid and the surface layer of the solid. The terms adsorption deals with the process in which molecules accumulate in the interfacial layer but desorption denotes the converse

process. The material in the adsorbed state is referred to as the ‗adsorbate‘, but that in the bulk gas or vapour phase prior to being adsorbed is called the ‗adsorptive‘. The penetration by the adsorbate molecules into the bulk solid phase is determined as

‗absorption‘. The term ‗sorption‘ is also used to denote both adsorption and absorption, when both occur simultaneously or cannot be distinguished (Dabrowski, 2003).Adsorption is recognized as the most efficient, promising and widely used fundamental approach in wastewater treatment process (Foo and Hameed, 2010)

## Adsorption Isotherm

This is a fundamental concept in the science of adsorption. It is the equilibrium relation between the quantity of the adsorbed material and the pressure or concentration in the bulk fluid phase at constant temperature. Apart from the result in the calorimetric measurements, the adsorption isotherm is the primary source of information in the adsorption process.

## Adsorption Isotherm Models

In general, an adsorption isotherm is an invaluable curve describing the phenomenon governing the retention (or release) or mobility of a substance from the aqueous porous media or aquatic environments to a solid-phase at a constant temperature and pH (Limousin*et al*., 2007). Adsorption equilibrium (the ratio between the adsorbed amount with the remaining in the solution) is established when an adsorbate containing phase has been contacted with the adsorbent for sufficient time, with its adsorbate concentration in the bulk solution is in a dynamic balance with the interface concentration (Dabrowski, 2003). Its physicochemical parameters together with the underlying thermodynamic assumptions provide an insight into the adsorption mechanism, surface properties as well as the degree of affinity of the adsorbents (Malek

and Farooq, 1996). Over the years, a wide variety of equilibrium isotherm models (Langmuir, Freundlich, Brunauer–Emmett–Teller, Redlich– Peterson, Dubinin– Radushkevich, Temkin, Toth, Koble–Corrigan, Sips, Khan, Hill, Flory–Huggins and Radke–Prausnitz isotherm), have been formulated in terms of three fundamental approaches (Bulut et al., 2008). Kinetic consideration is the first approach to be referred. Hereby, adsorption equilibrium is defined being a state of dynamic equilibrium, with both adsorption and desorption rates are equal (Langmuir, 1916).

## Two Major Isotherms Langmuir isotherm model

Langmuir adsorption isotherm, originally developed to describe gas–solid-phase adsorption onto activated carbon, has traditionallybeen used to quantify and contrast the performance of different bio-sorbents (Langmuir, 1916). In its formulation, this empirical model assumes monolayer adsorption (the adsorbed layer is one molecule in thickness), with adsorption can only occur at a finite (fixed) number of definite localized sites, that are identical and equivalent, with no lateral interaction and steric hindrance between the adsorbed molecules, even on adjacent sites (Padmesh et al., 2006). In its derivation, Langmuir isotherm refers to homogeneous adsorption, which each molecule possess constant enthalpies and sorption activation energy (all sites possess equal affinity for the adsorbate), with no transmigration of the adsorbatein the plane of the surface (Kundu and Gupta, 2006).

## Freundlich isotherm model

Freundlich isotherm is the earliest known relationship describing the non-ideal and reversible adsorption, not restricted to the formation of monolayer. This empirical model can be applied to multilayer adsorption, with non-uniform distribution of

adsorption heat and affinities over the heterogeneous surface. Historically, it is developed for the adsorption of animal charcoal, demonstrating that the ratio of the adsorbate onto a given mass of adsorbent to the solute was not a constant at different solution concentrations (Ahmaruzzaman, 2008). In this perspective, the amount adsorbed is the summation of adsorption on all sites (each having bond energy), with the stronger binding sites are occupied first, until adsorption energy are exponentially decreased upon the completion of adsorption process.

At present, Freundlich isotherm is widely applied in heterogeneous systems especially for organic compounds or highly interactive species on activated carbon and molecular sieves. The slope ranges between 0 and 1 is a measure of adsorption intensity or surface heterogeneity, becoming more heterogeneous as its value gets closer to zero. Whereas, a value below unity implies chemisorptions process where 1/n above one is an indicative of cooperative adsorption. Recently, Freundlich isotherm is criticized for its limitation of lacking a fundamental thermodynamic basis, not approaching the Henry‘s law at vanishing concentrations.

# CHAPTER THREE

## MATERIALS AND METHODS

The materials, instruments and experimental procedures for this research are presented in this chapter.

## List of materials and equipment

**Table 3.1. Laboratory equipment used for this research.**

|  |  |  |  |
| --- | --- | --- | --- |
| **S/N** | **MATERIALS/EQUIPMENT** | **MODEL** | **AVAILABILITY** |
| **1.** | Glass Beaker | BEX 230 | WREE Lab |
| **2.** | Digital pH meter | pHS-2S | WREE Lab |
| **3.** | Digital Electrical Conductivity  meter | Hanna HI 207 | WREE Lab |
| **4.** | Oven | Griffin 20X | WREE Lab |
| **5.** | 20ml sample bottles | PET 1 | WREE Lab |
| **6.** | Immersion Thermometer | NA | WREE Lab |
| **7.** | Weighing Balance | FA2004B | WREE Lab |
| **8.** | Incubator | Griffin TS | WREE Lab |
| **9.** | Reflux Machine | NA | WREE Lab |
| **10.** | Glass rod | Technico 200C | WREE Lab |
| **11.** | Lovibond Apparatus | S500C | WREE Lab |
| **12.** | Kjelahl Apparatus | NA | Soil Science Lab |
| **23.** | Masking Tape | NA | Samaru Market |
| **24.** | Foil Paper | NA | Samaru Market |
| **25.** | Marker (Parmanent Type) | Huaying 3200 | Samaru Market |
| **26.** | Rubber Clips | NA | Samaru Market |
| **27.** | Furnance | Gemco 20kW | IDC Lab |

## Table 3.2. Workshop Materials used during research

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| **S/N** | **MATERIALS/EQUIPMENT** | | | | **MODEL** | **AVAILABILITY** | |
| **1.** | 12.7 ball gauge | | | | P112FGP-WH | Samaru  market | Local |
| **2.** | 12.7 mm pipe | | | | P112FGP-WH | Samaru  market | Local |
| **3.** | 101.6 mm plug | | | | NA | Samaru  market | Local |
| **4.** | 101.6 mm pipe | | | | P112FGP-WH | Samaru  market | Local |
| **5.** | PVC gum | | | | AbroOATEY  31008 | Samaru  market | Local |
| **6.** | Silk Sieve | | | | NA | Samaru  market | Local |
| **7.** | Saw and Screw | | | | NA | Samaru  market | Local |
| **8.** | 12.7mm  washers | rubber | bolt | and | NA | Samaru market | Local |
| **9.** | 101.6 mm metal clip | | | | NA | Samaru  market | Local |
| **10.** | Pipe Threading Machine | | | | NA | ABU Works Dept. | |
| **11.** | Grinder | | | | NA |  | |
| **12.** | Weighing Scale | | | | SM-600 |  | |
| **23.** | 2mm and 4.7mm sieve | | | | ASTM 4.7/2 |  | |

**Table 3.3 Materials used for this study**

|  |  |  |  |
| --- | --- | --- | --- |
| **S/N** | **MATERIALS/EQUIPMENT** | **MODEL** | **AVAILABILITY** |
| **1.** | Sand (0.5mm to 0.7mm) | Rapid Sand | Samaru |
| **2.** | Wood derived biochar | NA | Samaru |

|  |  |  |  |
| --- | --- | --- | --- |
| **3.** | Greywater sample | NA | ABU, Suleiman Hostel |

## Experiment plan

* + 1. A known mas of 10kg of the trees (*Vitellariaparadoa, Khayasenegalenses, FicusSycomous)* was obtained, sundried for 10 days then oven dried at 105oC for three hours. The dried wood were then chipped into average sizes of 200mm each.
    2. The dried wood were then pyroluzed at 650oC using a Gemcofurnance.
    3. The resulting biochar was ground into different particle sizes and classified into (2mm <x<4.7mm and x<2mm)
    4. The classified biochar groups were then halved after which one set was activated using H2SO4 and then washed with distilled water.
    5. The resulting biochar and activated carbon were then used as filters in PVC constructed filter columns.
    6. Greywater from Ahmadu Bello University Suleiman Hostel was then passed through the filters at a hydraulic loading rate of 0.033m3/m2/day for 21 days.
    7. Their performance in remediating the greywater was analyzed using ANOVA.
    8. The effects of particle size, time, parent material and activation status between the sets were also considered during the remediation process.

## Preparation of filter material.

* + 1. **Preparation of Biochar**

The trees used for this study: from *Vitellariaparadoa* (Shea tree), *Khayasenegalenses* (Mahogany)and *FicusSycomous*(Fig) were identified in Samaru area of Zaria in Nigeria and its stems were cut down. The stems were then chopped down into smaller pieces using an axe and 10 kg of each of the tree were obtained.

The 50kg obtained from each tree was sun dried for 48 hours to reduce the moisture content. They were further dried at a temperature of 105oC using a furnace for three hours.

The dried chipped wood was then pyrolyzed at a temperature of 650oC using a 20kWGemco furnace and the resulting biochar were collected, grinded and sieved using a 4.7mm standard sieve and a 2mm standard sieve. This resulted to obtaining two sets of biochar for each tree (one with a range from 2mm to 4.7mm and the other set of less than 2mm). Each set was then halved resulting to a total of 12 sets. (four for each parent material). The first half of the set (a total of 6 samples, 2 for parent material) were then activated with concentrated H2SO4.

During the pyrolysis of the woods, same quantity of 50 kg each were subjected to the furnace for pyrolysis but their yield were different. Their yield obtained after the pyrolysis was proportional to their lignin content. The shea-tree had the most yield of 36% after the pyrolysis with mahoganyand the fig yielding 30% and 25% respectively.

## Activation of Biochar

A total of 6 samples (fine and coarse) from each tree were then soaked in a solution ofSulphuric acid diluted to 10% with distilled water for 24 hours after which it was dried and charred. The resulting activated biochar was then washed using distilled water and sun dried for 10 hours.

## Construction of Filter Columns

The filter columns were constructed using 101.6mm diameter Polyvinyl Chloride (PVC) pipes, 12.7mm diameter PVC pipes, stoppers, control valves, PVC gum and sets of bolts and washers. 400 mm of the 101.6mm diameter PVC pipe was measured and cut. A stopper was then fitted at the bottom of the 101.6mm diameter pipe using a pvc gum. A hole of half 12.7mm diameter was drilled at the center of the stopper using a hand drill and a pipe of length 50mm and 12.7mm diameter with a threaded edge was then inserted in the hole and held firm by12.7mmbolts and washers. The edge of the 12.7mmdiameter pipe was then fitted with a control valve.

Figure 3.1. Schematic Diagram of filter column

## Preparation and Loading of Filter Columns

The top of the12.7mm diameter pipe inside the filter was sealed with a 212µm mesh. A filter membrane made from cotton which measures about 20mm thick was then added to the 212µm mesh surface. Rapid sand of sizes between 0.5 and 0.7mm was washed and poured in to a level of 5cm from inside the column. Seven hundred grams (700g) of each of the biochar and activated Carbon were then measured and added to their specific column which was then labelled appropriately. The set biochar column gave a total of Twelve (12) columns each with different biochar content with a thirteenth column with no biochar and sand in it which served as the control column.

15 litres of greywater from Suleiman Hostel was obtained at 8 AM in the morning and another 15 litres at 4 PM in the evening was obtained and mixed to obtain a composite sample. 2 litres of the composite sample was then loaded in each of the 13 columns.200ml of the filtrate was collected and replaced daily with composite sample of that day.

## Determination of Lignin Content of the filter media

A fraction (0.3)g of dried extracted raw biomass was weighed in glass test tubes and 3 mL of 72% H2SO4 was added. The sample was kept at room temperature for 2 hours with carefully shaking at 30 min intervals to allow for complete hydrolysis. After the initial hydrolysis, 84 ml of distilled water was added. The second step of hydrolysis was made to occur in an autoclave for 1 h at 121°C. The slurry was then cooled at room temperature. Hydrolyzates were filtered through vaccum using a filtering crucible. The acid insoluble lignin was determined by drying the residues at 105oC and accounting for ash by incinerating the hydrolyzed samples at 575 oC in a muffle furnace. The acid soluble lignin fraction was determined by measuring the absorbance of the acid

hydrolyzed samples at 320 nm. The lignin content was calculated as the summation of acid insoluble lignin and acid soluble lignin. This method was suggested by United State Department of Energy, Sluiter*et al* (2008).

## Determination of Physical Parameters

The physical properties of the filters which include bulk density, particle density, porosity, air dry water content were determined using Standard Soil Physics procedures as reported by Hillel (1982). Particle size range was determined by using 4.7mm and 2mm standard sieves while pyrolysis yield was determined percentage wise.

## Laboratory Analysis of Samples

Daily samples obtained from the 200ml bottles were tested in the laboratory immediately. Physico-Chemical parameters of pH, EC, Tot-N, Tot-P, PO4-P, NO3-N, NH3-N, BOD and COD were tested and recorded for a period of 21 days.pH and Electrical Conductivity (EC) were determined using a portable standardized Hanna instrument.

Electrical conductivity were tested using a standardized calibrated instrument while the other parameters were analyzed using laboratory methods. The table below gives the summary of all the laboratory analysis methods, standards, equipment and reagents used during the analysis. Total Nitrogen was determined by standard Kjaldahl method after dilution with Bromocresol given as indicator. Nitrate Nitrogen was also determined with the Kjaldahl method using Magnesium Oxide and Boric Acid with Devarda‘s alloy. Ammonium Nitrogen was analyzed using the Lovibond Apparatus with Nessler reagent and Rochelle Salt. The Chemical Oxygen Demand (COD) was determined using the Potassium Dichromate method. On the other hand, BOD was determined after incubating for five (5) days using alkali iodized-azadide reagent,

Manganese Sulphate and Sulphuric acid using starch as an indicator on the titration with Sodiumthiosulphate. The yellow method was used to determine available Phosphorus method with a spectrophotometer using Ascorbic Acid, Potassium, Ammonium vanadate, Ammonium molybdate, nitric acid and Potassium antimonyl tartrate. Similarly, Total Phosphorus was determined with the spectrophotometer using Ammonium molybdate, Hydrochloric Acid and Tin Chloride. Table I shows a summary of procedures and instruments used.

Treatment Efficiencies were calculated using the formular

𝐸 = 𝐶𝑖𝑛 −𝐶𝑜𝑢𝑡

𝐶𝑖𝑛

(3.1)

Where E is efficiency in %

Cin is initial concentration of pollutants Cout is Final concentration of pollutants

Table 3.4 Laboratory Procedures and reagents used

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **PARAMETER** | **REGENTS** | **MEASUREMENT RANGE** | **UNIT** | **METHOD** | **APPARATUS** |
| **COD** | H2SO4, K2CR2O7, Ferrous  indicator, Ferrous AmoniumSulphate | Not Applicable | mgl-1 | Quantitative | - |
| **BOD** | MaganeseSulphate, Alkali iodized-azadide reagent,  H2SO4, Na2S2O3, starch | Not Applicable | mgl-1 | Winkler | - |
| **NH4-N** | Nesslers reagent, Rochelle salt | COLOUR INTENSITY | mgl-1 | Colorimetric | lovibond |
| **NO3-N** | Boric acid, Magnesium  Oxide, Devarda‘s alloy | Not Applicable | mgl-1 | Kjeldahl | Kjeldahl Apparatus |
| **TOT-N** | H2SO4, NaOH, Boric SoiumSulphate,  CupperSulphate, Bromocresol Green Inicator | Not Applicable | mgl-1 | Kjeldahl | Kjeldahl Apparatus |
| **PO4-P** | Ascorbic acid, potassium antimonyl tartrate, amonium  vanadate, ammonium molybate, nitric acid | 420nm | mgl-1 | Yellow Method | Spectrophotometer |
| **TOT-P** | Ammonium molybdate, , Hydrochloric Acid, Tin  Chloride | 500 – 700 nm | mgl-1 |  | Spectrophotometer |
| **pH** | NONE | 0 – 14 | NONE | Direct | Hanna Instrument |
| **EC** | NONE | 0 – 20,000 | µS/cm | Direct | Hanna Instrument |
| **Lignin** | H2SO4 | 320nm | %w/w | Indirect | Spectrophotometer |

## CHAPTER FOUR RESULTS AND DISCUSSION

* 1. **Results**

The 200ml sample collected daily from all the thirteen filters were tested in the laboratory and values were recorded throughout the 21 days of the experiment. The experiment was replicated three times and the results were analyzed using Microsoft Excel 2013 and Instat Statistic Package Version 3 (Graph-pad). Results for each analyzed parameter is presented in a pair of tables.

The weekly summary of the adsorbed concentration of the pollutants is presented as Mean+ SEM (Standard error of mean). Results presented across the columns shows comparisons between the three parent materials and also comparison between Biochar and Activated Carbon while results presented down the rows showcomparisons between the different particle sizes and different time (in weeks). Differences between the columns are calculated using Fisher Statistics values and Critical Fisher Values (FC) at minimum of 95% confidence limit.

The comparisons between particle sizes and activation status is shown in terms of P values. P values greater than 0.05 were considered Not Significant and labelled (NS) while P values less than 0.05 and less than 0.01 were considered significant and Highly significant and are represented with asterix signs (\*\*) and (\*) respectively. The P-value comparisons were calculated analyzed Bonferroni Multiple Comparison Test .

Table 4.1 shows all the thirteen columns and their labels with their corresponding parent material, activation status and particle sizes.

Table 4.2 shows the calculated physical properties of the filters, their calculated lignin content and their percentage yield after pyrolysis.

TABLE 4.1 DESCRIPTION OF COLUMNS

|  |  |  |  |
| --- | --- | --- | --- |
| **COLUMNS** | **FILTER MATERIAL** | **PARTICLE SIZE**  **(mm)** | **FILTER TYPE** |
| COLUMN 1 | FIG | 2 – 4.7 | BIOCHAR |
| COLUMN 2 | FIG | < 2 | BIOCHAR |
| COLUMN 3 | MAHOGANY | 2 – 4.7 | BIOCHAR |
| COLUMN 4 | MAHOGANY | < 2 | BIOCHAR |
| COLUMN 5 | SHEA TREE | 2 – 4.7 | BIOCHAR |
| COLUMN 6 | SHEA TREE | < 2 | BIOCHAR |
| COLUMN 7 | FIG | 2 – 4.7 | ACTIVATED CARBON |
| COLUMN 8 | FIG | < 2 | ACTIVATED CARBON |
| COLUMN 9 | MAHOGANY | 2 – 4.7 | ACTIVATED CARBON |
| COLUMN 10 | MAHOGANY | < 2 | ACTIVATED CARBON |
| COLUMN 11 | SHEA TREE | 2 – 4.7 | ACTIVATED CARBON |
| COLUMN 12 | SHEA TREE | < 2 | ACTIVATED CARBON |
| COLUMN CONTROL | **-** | **-** | **-** |

## Table 4.2 Physical properties of the parent material used.

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| **Parameter** | **Fig tree** | | **Mahogany** | | **Shea tree** | |
| **Particle size (mm)** | 2 – 4.7 | <2 | 2 – 4.7 | <2 | 2 – 4.7 | <2 |
| **Air dry water content (%)** | 61 | 61 | 65 | 65 | 63 | 63 |
| **3**  **Bulk Density (kg/m )** | 307 | 296 | 283 | 270 | 296 | 261 |
| **3**  **Particle Density (kg/m )** | 1694 | 1694 | 1810 | 1810 | 1940 | 1940 |
| **Lignin Content (%w/w)** | 21.4 | 21.4 | 28.9 | 28.9 | 34.9 | 34.9 |
| **Yield after Pyrolysis (%)** | 25 | 25 | 30 | 30 | 36 | 36 |
| **Porosity (%)** | 83 | 71 | 74 | 61 | 79 | 65 |

Table 4.3 shows adsorbed Total Phosphorus from each filters presented as Mean±Standard Error of Mean. Compared Adsorbed concentration of Total Phosphorus for each parent material is presented across the columns while the comparison factors of Time and Particle size is presented down the rows. The results shows averagely same performance for each of the filters. Although minor differences can be observed, the differences does not qualify as statistically significant.

The table also shows that Total Phosphorus adsorption occurred higher in the first week with slight reduction in the second and third week.

## Table 4.3 Adsorbed Concentration of Total Phosphorus From Filters

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Time (Weeks)** | **PS**  **(mm)** |  | **Biochar (mgl-1)** |  | **Activated Carbon (mgl-1)** | |  | **Fisher values** | |
| **Fig** | **Mahogany** | **Shea** | **Fig** | **Mahogany** | **Shea** | **FV** | **FC**  **∞=5%** |
| **1** | 2 – 4 | 48.414±6.462 | 47.277±6.514 | 47.494±6.516 | 43.677±6.663 | 40.620±5.623 | 41.169±6.571 | 0.146 | 1.906 |
|  | < 2 | 47.663±6.646 | 47.611±6.508 | 47.361±6.676 | 37.386±6.512 | 37.390±6.251 | 40.641±6.737 |  |  |
| **2** | 2 – 4 | 37.191±0.633 | 36.697±0.565 | 36.497±0.536 | 30.677±0.639 | 30.670±0.604 | 29.861±0.489 | 50.579 | 1.906 |
|  | < 2 | 36.756±0.687 | 36.247±0.647 | 36.043±0.589 | 27.913±0.484 | 27.369±0.505 | 27.406±0.520 |  |  |
| **3** | 2 – 4 | 26.929±0.998 | 26.286±1.022 | 25.747±1.065 | 21.434±0.885 | 21.060±0.985 | 21.181±0.768 | 10.923 | 1.906 |
|  | < 2 | 26.163±1.052 | 24.111±1.163 | 23.813±1.142 | 17.896±0.985 | 17.970±0.840 | 20.144±0.666 |  |  |

Table 4.4 shows comparisons between Biochar Filter Columns and Activated Carbon Filter Columns. It also shows comparisons between filters with less than 2mm particle size and filters with a range of 2mm to 4.7mm particle size. Using P values as a standard for the comparisons, the compared results showed no significant difference between all columns compared

## Table 4.4 Level of Significant differencesand Comparisons for Total Phosphorus

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Time**  **(weeks)** | **Filter**  **Media** | **Particle**  **Size Comparison.** | **P**  **Value** | **Activation status**  **Comparison** | **P**  **Value** |
| **1** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
| **2** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
| **3** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |

Table 4.5 shows results for adsorbed COD presented as Mean±Standard Error of Mean. It presents adsorbed results from different parent materials and activation status across the columns while results for different time (in weeks ) and different particle sizes are presented down the rows. Differences between the columns are shown using Fisher values and Critical Fisher values are 95% confidence limit.

The table shows that there were no observed significant differences between all the compared parent materials in terms of COD adsorption. It however shows that more adsorption was observed in the second week with less in the first and third week respectively.

## Table 4.5 Adsorbed Concentration of COD From Filters

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **TIME**  **(Weeks)** | **PS**  **(mm)** |  | **Biochar (mgl-1)** |  | **Activated Carbon (mgl-1)** | |  | **Fisher values** | |
| **Fig** | **Mahogany** | **Shea** | **Fig** | **Mahogany** | **Shea** | **FV** | **FC**  **∞=5%** |
| **1** | 2 – 4 | 257.14±24.247 | 248.57±34.235 | 364.29±28.356 | 214.29±30.147 | 281.43±27.276 | 335.71±35.245 | 3.435 | 1.906 |
|  | < 2 | 245.71±25.435 | 262.86±33.645 | 364.29±26.624 | 211.43±28.320 | 298.57±26.675 | 348.57±35.752 |  |  |
| **2** | 2 – 4 | 312.86±42.410 | 298.57±44.477 | 351.43±43.008 | 218.57±39.123 | 294.29±39.088 | 305.71±38.350 | 0.946 | 1.906 |
|  | < 2 | 302.86±46.080 | 289.57±46.723 | 345.71±44.873 | 214.29±39.088 | 292.86±42.802 | 314.29±44.178 |  |  |
| **3** | 2 – 4 | 181.43±7.997 | 171.43±7.997 | 218.57±10.335 | 125.71±8.411 | 188.57±6.335 | 187.14±8.650 | 9.554 | 1.906 |
|  | < 2 | 160.60±10.690 | 157.14±11,488 | 205.57±12.509 | 118.57±9.368 | 172.86±7.781 | 177.14±9.440 |  |  |

Table 4.6 shows comparisons between Biochar and Activated Carbon and also between

<2mm particle sized filters and 2 – 4 mm particle sized filters. Using P value as the factor for the comparisons, there were no observed significant differences between all made comparisons except for a single case of the Fig derived filters in week one were comparisons between Biochar and Activated Carbon showed a P value of <0.001 (\*\*) and

<0.05(\*) respectively.

## Table4.6 Level of Significant difference and Comparisons for COD

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Time**  **(weeks)** | **Filter**  **Media** | **Particle**  **Size comparison.** | **P**  **Value** | **Activation status**  **comparison** | **P**  **Value** |
| **1** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \* |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
| **2** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
| **3** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |

Table 4.7 shows adsorbed Nitrate Nitrogen (NO3-N) presented as Mean±Standard Error of mean. It shows comparisons between different parent materials across the columns while comparisons for different particle sizes and different time (in weeks) are presented down the rows.

The table indicates that there were no observed significant differences between the different particle sizes, different parent materials and differences in activation status. There was however observed differences between concentrations adsorbed in the first, second and third week with the rate of adsorption more in the second week.

## Table4.7 Adsorbed Concentration of NO3-N from filters

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Time (Weeks)** | **Ps (mm)** |  | **Biochar (mgl-1)** |  | **Activated Carbon (mgl-1)** | |  | **Fisher Values** | |
| **Fig** | **Mahogany** | **Shea** | **Fig** | **Mahogany** | **Shea** | **FV** | **FC**  **∞=5%** |
| **1** | 2 – 4 | 77.214±9.598 | 81.286±7.724 | 82.214±9.461 | 73.286±5.410 | 78.429±8.879 | 74.286±5.524 | 0.329 | 1.906 |
|  | < 2 | 85.714±8.937 | 83.857±8.374 | 84.286±8.420 | 71.286±8.959 | 77.357±6.955 | 81.357±7.601 |  |  |
| **2** | 2 – 4 | 31.286±4.276 | 33.071±5.085 | 32.500±5.032 | 33.000±4.795 | 29.143±3.892 | 33.080±5.098 | 0.064 | 1.906 |
|  | < 2 | 33.08±5.369 | 33.500±5.032 | 32.000±4.923 | 33.000±4.795 | 32.500±4.689 | 37.500±5.032 |  |  |
| **3** | 2 – 4 | 14.000±1.328 | 16.000±1.291 | 15.500±1.041 | 12.500±1.848 | 11.500±2.380 | 13.429±1.605 | 0.974 | 1.906 |
|  | < 2 | 15.500±1.041 | 16.000±1.291 | 15.000±1.826 | 12.500±1.500 | 13.070±1.826 | 12.000±2.141 |  |  |

Table 4.8 shows comparisons of adsorbed NO3-N between Biochar and Activated Carbon and also between <2mm particle sized filters and 2 – 4 mm particle sized filters. Using P value as the factor for the comparisons, there were not observed significant differences between all comparisons.

## Table 4.8 Level of significant differences and comparisons for NO3-N.

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Time**  **(weeks)** | **Filter**  **Media** | **Particle**  **Size Comparison.** | **P**  **Value** | **Activation Status**  **Comparison** | **P**  **Value** |
| **1** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
| **2** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
| **3** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |

Table 4.9 shows results for adsorbed Phosphate Phosphorus (PO4-P) presented as Mean±Standard Error of Mean. It presents adsorbed results from different parent materials and activation status across the columns while results for different time (in weeks) and different particle sizes are presented down the rows. Differences between the columns are shown using Fisher values and Critical Fisher values are 95% confidence limit.

The table also showed that there were no observed significant differences between all the compared parent materials in terms of PO4-P adsorption. It however shows that more adsorption was observed in the first week with less in the second and third week respectively.

## Table 4.9 Adsorbed Concentration of PO4-PFrom Filters

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Time (Weeks)** | **PS**  **(mm)** |  | **Biochar (mgl-1)** |  | **Activated Carbon (mgl-1)** | |  | Fisher Values | |
| **Fig** | **Mahogany** | **Shea** | **Fig** | **Mahogany** | **Shea** | **FV** | **FC**  **∞=5%** |
| **1** | 2 – 4 | 148.33±19.817 | 144.69±19.975 | 145.23±19.981 | 133.81±20.431 | 124.84±17.213 | 126.16±20.145 | 0.414 | 1.906 |
|  | < 2 | 146.33±19.823 | 145.87±20.141 | 145.17±20.459 | 115.92±19.966 | 114.56±19.166 | 124.52±20.663 |  |  |
| **2** | 2 – 4 | 113.99±1.941 | 112.47±1.733 | 111.86±1.644 | 94.027±1.959 | 94.317±2.128 | 91.524±1.499 | 51.095 | 1.906 |
|  | < 2 | 112.47±2.095 | 111.18±1.986 | 110.47±1.806 | 84.409±1.451 | 83.893±1.542 | 83.321±1.329 |  |  |
| **3** | 2 – 4 | 82.539±3.061 | 80.553±3.132 | 78.917±3.264 | 65.710±2.720 | 64.553±3.019 | 64.926±2.356 | 10.848 | 1.906 |
|  | < 2 | 80.191±3.224 | 73.903±3.566 | 72984±3.496 | 54.844±2.954 | 55.077±2.576 | 62.030±2.114 |  |  |

Table 4.10 shows comparisons of adsorbed PO4-P between Biochar and Activated Carbon and also between <2mm particle sized filters and 2 – 4 mm particle sized filters. Using P value as the factor for the comparisons, there were not observed significant differences between columns of different particle sizes. Significant differences were observed between biochar and activated Carbon in the second and third week

## Table 4.10 Level of Significant Difference and Comparison ForPO4-P

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Time**  **(weeks)** | **Filter**  **Media** | **Particle**  **Size Comparison.** | **P**  **Value** | **Activation status**  **Comparison** | **P**  **Value** |
| **1** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
| **2** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
| **3** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | \* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |

Table 4.11 shows results for adsorbed Total Nitrogen (TOT-N) presented as Mean±Standard Error of Mean. It presents adsorbed results from different parent materials and activation status across the columns while results for different time (in weeks) and different particle sizes are presented down the rows. Differences between the columns are shown using Fisher values and Critical Fisher values are 95% confidence limit.

The table shows that there were no observed significant differences between all the compared parent materials in terms of Total Nitrogen adsorption. It however shows that more adsorption was observed in the first week with less in the second and third week respectively.

## Table4.11 Adsorbed Concentration of Total NitrogenFrom Filters

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Time (weeks)** | **PS**  **(mm)** |  | **Biochar (mgl-1)** |  | **Activated carbon (mgl-1)** | |  | **Fisher Values** | |
| **Fig** | **Mahogany** | **Shea** | **Fig** | **Mahogany** | **Shea** | **FV** | **FC**  **∞=10%** |
| **1** | 2 – 4 | 22.000±1.826 | 23.000±2.000 | 23.000±2.000 | 20.00±1.826 | 23.00±2.000 | 23.000±2.000 | 0.225 |  |
|  | < 2 | 23.000±2.000 | 23.000±2.000 | 21.00±2.160 | 22.000±1.826 | 23.000±2.000 | 23.000±2.000 |  |  |
| **2** | 2 – 4 | 12.071±3.299 | 11.071±2.985 | 11.071±2.985 | 12.071±3.299 | 13.071±3.855 | 13.071±3.855 | 0.046 |  |
|  | < 2 | 13.071±3.855 | 12.071±3.299 | 12.071±3.299 | 12.071±3.299 | 13.071±3.855 | 13.071±3.855 |  |  |
| **3** | 2 – 4 | 6.214±0.706 | 6.214±0.706 | 6.214±0.706 | 7.214±1.327 | 6.714±0.938 | 6.714±1.401 | 0.172 |  |
|  | < 2 | 6.714±0.937 | 6.214±0.706 | 5.714±0.786 | 7.214±1.327 | 6.714±1.401 | 6.714±1.401 |  |  |

Table 4.12 shows comparisons of adsorbed Total Nitrogen between Biochar and Activated Carbon and also between <2mm particle sized filters and 2 – 4 mm particle sized filters. Using P value as the factor for the comparisons, there were not observed significant differences between all comparisons.

## Table 4.12 Level of Significant Difference and Comparisons for Total Nitrogen

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Time**  **(weeks)** | **Filter**  **Media** | **Particle**  **Size Comparison.** | **P**  **Value** | **Activation status**  **Comparison** | **P**  **Value** |
| **1** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
| **2** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
| **3** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |

Table 4.13 shows results for adsorbed BOD presented as Mean±Standard Error of Mean. It presents adsorbed results from different parent materials and activation status across the columns while results for different time (in weeks) and different particle sizes are presented down the rows. Differences between the columns are shown using Fisher values and Critical Fisher values at 95% confidence limit.

The table shows that there were no observed significant differences between all the compared parent materials in terms of BOD adsorption. It also shows that the rate of BOD reduction in the filters increase with time unlike other parameters.

## Table 4.13 Adsorbed Concentration Of BOD From Filters

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Time (weeks)** | **PS**  **(mm)** |  | **Biochar (mgl-1)** |  | **Activated carbon (mgl-1)** | |  | **Fisher values** | |
| **Fig** | **Mahogany** | **Shea** | **Fig** | **Mahogany** | **Shea** | **Fv** | **Fc**  **∞=5%** |
| **1** | 2 – 4 | 258.00±25.962 | 156.00±18.601 | 212.00±26.533 | 242.00±25.573 | 226.00±22.716 | 214.00±21.119 | 1.421 | 1.998 |
|  | < 2 | 250.00±26.077 | 180.00±36.742 | 238.00±23.324 | 244.00±23.791 | 226.00±22.716 | 210.00±20.736 |  |  |
| **2** | 2 – 4 | 274.00±10.296 | 220.00±10.000 | 238.00±13.928 | 256.00±10.296 | 250.00±11.402 | 228.00±11.576 | 2.428 | 1.998 |
|  | < 2 | 268.00±8.602 | 223.00±9.695 | 240.00±12.247 | 250.00±8.944 | 244.00±11.225 | 276.80±11.576 |  |  |
| **3** | 2 – 4 | 310.00±16.733 | 278.00±16.248 | 270.00±16.125 | 296.800±16.852 | 296.00±11.662 | 276.80±11.662 | 1.267 | 1.998 |
|  | < 2 | 302.04±16.533 | 272.00±16.533 | 268.00±8.00 | 272.00±9.165 | 292.000±11.576 | 260.00±13.416 |  |  |

Table 4.14 shows comparisons of adsorbed BOD between Biochar and Activated Carbon and also between <2mm particle sized filters and 2 – 4 mm particle sized filters. Using P value as the factor for the comparisons, there were no observed significant differences between all comparisons

## Table 4.14 Level of significant Difference and comparisons for BOD Results

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Time**  **(weeks)** | **Filter**  **Media** | **Particle**  **Size comparison.** | **P**  **Value** | **Activation status**  **comparison** | **P**  **Value** |
| **1** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
| **2** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
| **3** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |

Table 4.15 shows results for adsorbed EC presented as Mean±Standard Error of Mean. It presents adsorbed results from different parent materials and activation status across the columns while results for different time (in weeks) and different particle sizes are presented down the rows. Differences between the columns are shown using Fisher values and Critical Fisher values at 95% confidence limit.

The table shows that there were no observed significant differences between all the compared parent materials in terms of EC adsorption. It also shows that the rate of EC reduction in the filters reduces with time,

## Table 4.15 Adsorbed Concentration of ECFrom Filters

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Time (Weeks)** | **PS**  **(mm)** | **Biochar (µS/CM)** | |  | **Activated Carbon (µS/cm)** | |  | **Fisher Values** | |
| **Fig** | **Mahogany** | **Shea** | **Fig** | **Mahogany** | **Shea** | **FV** | **FC**  **∞=5%** |
| **1** | 2 – 4 | 3279.6±210.38 | 2044.1±257.00 | 2951.9±478.15 | 4280.6±138.36 | 4660.1±177.33 | 4495.1±174.59 | 18.322 | 1.906 |
|  | < 2 | 2786.6±358.68 | 1609.7±326.11 | 2484.0±167.66 | 3966.3±141.91 | 4534.0±175.13 | 4422.37±137.84 |  |  |
| **2** | 2 – 4 | 2638.6±119.67 | 2284.0±13.968 | 2844.0±90.563 | 3696.3±130.15 | 3905.1±148.14 | 3827.0±154.70 | 55.379 | 1.906 |
|  | < 2 | 1222.9±119.99 | 1330.6±71.742 | 2320.9±112.49 | 3523.6±162.25 | 3858.1±149.28 | 3805.6157.30 |  |  |
| **3** | 2 – 4 | 2588.6±158.58 | 2239.1±61.283 | 2393.4±39.308 | 2991.9±100.50 | 3098.1±10.343 | 3021.1±103.91 | 59.090 | 1.906 |
|  | < 2 | 1080.3±43.269 | 1224.4±81.236 | 2002.1±38.629 | 2846.9±138.36 | 3079.7±104.86 | 2966.9±105.17 |  |  |

Table 4.16 shows comparisons of adsorbed EC between Biochar and Activated Carbon and also between <2mm particle sized filters and 2 – 4 mm particle sized filters. Using P value as the factor for the comparisons, it was observed that there were no significant differences between the compared particle sizes. All comparisons between Biochar and Activated Carbon however showed significant differences.

## Table 4.16 Level of Significant Differences and Comparisons for EC

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Time**  **(weeks)** | **Filter**  **Media** | **Particle**  **Size comparison.** | **P**  **Value** | **Activation status**  **comparison** | **P**  **Value** |
| **1** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
| **2** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
| **3** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |

Table 4.17 shows results for adsorbed NH4-N presented as Mean±Standard Error of Mean. It presents adsorbed results from different parent materials and activation status across the columns while results for different time (in weeks) and different particle sizes are presented down the rows. Differences between the columns are shown using Fisher values and Critical Fisher values at 95% confidence limit.

The table shows that there were no observed significant differences between all the compared parent materials in terms of NH4-N adsorption. It also shows that the rate of NH4-N reduction in the filters reduces with time.

## Table 4.17 Adsorbed Concentration of NH4-NFrom Filters

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Time (weeks)** | **PS**  **(mm)** |  | **Biochar (mgl-1)** |  | **Activated carbon (mgl-1)** | |  | **Fisher values** | |
| **Fig** | **Mahogany** | **Shea** | **Fig** | **Mahogany** | **Shea** | **Fv** | **Fc**  **∞=5%** |
| **1** | 2 – 4 | 1.294±0.062 | 1.283±0.047 | 1.289±0.043 | 1.226±0.082 | 1.266±0.046 | 1.420±0.056 | 4.681 | 1.906 |
|  | < 2 | 1.117±0.058 | 1.329±0.057 | 1.220±0.036 | 1.346±0.075 | 1.540±0.074 | 1.923±0.045 |  |  |
| **2** | 2 – 4 | 0.302±0.214 | 0.537±0.170 | 0.263±0.139 | 0.754±0.103 | 0.800±0.099 | 0.897±0.094 | 3.192 | 1.906 |
|  | < 2 | 0.317±0.166 | 0.417±0.176 | 0.789±0.895 | 0.777±0.094 | 0.931±0.688 | 0.729±0.082 |  |  |
| **3** | 2 – 4 | -0.062±0.145 | 0.074±0.131 | 0.143±0.151 | 0.617±0.143 | 0.651±0.128 | 0.754±0.153 | 5.366 | 1.906 |
|  | < 2 | -0.058±0.140 | 0.143±0.134 | 0.880±0.147 | 0.817±0.143 | 0.766±0.140 | 0.423±0.317 |  |  |

Table 4.18 shows comparisons of adsorbed NH4-N between Biochar and Activated Carbon and also between <2mm particle sized filters and 2 – 4 mm particle sized filters. Using P value as the factor for the comparisons, three comparisons showed significant differences in the particle size comparisons. Similarly, three comparisons showed significant differences in the Activation Status comparison as shown in the table.

## Table 4.18 Level of Significant Difference and Comparisons for NH4-N

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Time**  **(weeks)** | **Filter**  **Media** | **Particle**  **Size comparison.** | **P**  **Value** | **Activation status**  **comparison** | **P**  **Value** |
| **1** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \* |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | \*\* | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | \*\* | B<2 VS AC<2 | NS |
| **2** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
| **3** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | \* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | \* | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |

Table 4.19 showed results for pH for all the filter columns. Results comparisons for different parent materials and activation status are read across columns while comparisons for different particle sizes and different time are shown down the rows.

The table showed that there were no observed significant differences between the different parent materials. It also shows that there were significant differences between the Biochar and Activated Carbon.

## Table 4.19 Direct Reading of pHFrom Filters

|  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Time (weeks)** | **PS**  **(mm)** |  | **Biochar** |  | **Activated carbon** | |  | **Fisher values** | |
| **Fig** | **Mahogany** | **Shea** | **Fig** | **Mahogany** | **Shea** | **Fv** | **Fc**  **∞=5%** |
| **1** | 2 – 4 | 7.674±0.171 | 8.043±0.092 | 8.780±0.065 | 6.829±0.052 | 6.800±0.148 | 6.90±0.135 | 45.32 | 1.641 |
|  | < 2 | 8.064±0.076 | 8.43±0.250 | 8.84±0.142 | 6.757±0.104 | 6.90±0.122 | 6.883±0.135 |  |  |
| **2** | 2 – 4 | 7.211±0.208 | 7.494±0.159 | 7.820±0.152 | 6.909±0.284 | 6.874±0.202 | 6.830±0.184 | 5.38 | 1.641 |
|  | < 2 | 8.114±0.351 | 7.830±0.195 | 8.253±0.261 | 6.993±0.201 | 6.773±0.205 | 7.019±0.156 |  |  |
| **3** | 2 – 4 | 7.596±0.145 | 7.449±0.071 | 7.396±0.086 | 7.283±0.208 | 7.177±0.168 | 7.034±0.194 | 7.062 | 1.641 |
|  | < 2 | 7.726±0.086 | 7.507±0.146 | 8.234±0.148 | 7.203±0.201 | 7.196±0.193 | 7.236±0.147 |  |  |

Table 4.20 shows comparisons of pH between Biochar and Activated Carbon and also between <2mm particle sized filters and 2 – 4 mm particle sized filters. Using P value as the factor for the comparisons, there were no observed significant differences between all the compared particle sizes. All the comparisons between Biochar and Activated Carbon showed significant difference in the first week. The columns made from Mahogany and Shea showed significant differences in the second column while only the <2mm Shea columns showed significant difference in the third week.

## Table 4.20 Level of Significant Difference and Comparisons for pH

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Time**  **(weeks)** | **Filter**  **Media** | **Particle**  **Size comparison.** | **P**  **Value** | **Activation status**  **comparison** | **P**  **Value** |
| **1** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
| **2** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | \*\* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \* |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | \* |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |
| **3** | Fig | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC2 – 4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Mahogany | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | NS |
|  | Shea | B2 – 4.7 VS B<2 | NS | B2 – 4.7 VS AC<4.7 | NS |
|  |  | AC2 – 4.7VS AC<2 | NS | B<2 VS AC<2 | \*\* |

Table 4.21 shows the measured level of pollutants at the end of the first week. All results are reported in MGL-1 except for pH which has no unit and EC which is reported in µS/cm.

## Table 4.21 Summary of Measured Parameters for All Columns in Week 1

|  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Parameter** | **Initial**  **Reading** |  |  | **Biochar** | |  |  |  |  | **Activated carbon** | |  |  |
|  | (Control) | FIG  (2-.7MM) | FIG  (<2MM) | MAH  (2-.7MM) | MAH  (<2MM) | SHEA  (2-.7MM) | SHEA  (<2MM) | FIG  (2-.7MM) | FIG  (<2MM) | MAH  (2-.7MM) | MAH  (<2MM) | SHEA  (2-.7MM) | SHEA  (<2MM) |
| TOT – P | 90 | 1.21 | 1.64 | 2.61 | 2.64 | 2.34 | 2.72 | 7.31 | 11.26 | 8.24 | 11.87 | 8.64 | 10.72 |
| COD | 600 | 40 | 50 | 60 | 60 | 20 | 10 | 150 | 150 | 80 | 60 | 60 | 50 |
| NO3-N | 125 | 14 | 7 | 7 | 7 | 10.5 | 7 | 7 | 7 | 17.5 | 7 | 7 | 7 |
| pH | 8.5 | 7.1 | 7.2 | 7.2 | 7.3 | 7.3 | 7.41 | 8.12 | 8.14 | 8.0 | 8.7 | 8.51 | 9.13 |
| PO4-P | 275.85 | 3.71 | 5.03 | 8.0 | 8.09 | 7.17 | 8.34 | 22.41 | 34.51 | 25.56 | 36.38 | 26.48 | 32.86 |
| TOT-N | 35 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 |
| BOD | 400 | 50 | 60 | 110 | 190 | 90 | 60 | 60 | 60 | 70 | 70 | 80 | 80 |
| EC | 7231 | 3161 | 4017 | 4132 | 4712 | 274 | 3192 | 1689 | 1804 | 1301 | 1370 | 1414 | 1368 |
| NH4-N | 1.9 | 0.44 | 0.72 | 0.52 | 0.52 | 0.64 | 0.52 | 0.64 | 0.64 | 0.56 | 0.52 | 0.64 | 0.8 |

Table 4.21 shows the measured level of pollutants at the end of the first week. All results are reported in MGL-1 except for pH which has no unit and EC which is reported in µS/cm.

## Table 4.22 Summary of Measured ParametersFor All Columns In Week 2

|  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Parameter** | **Initial**  **Reading** |  |  | **Biochar** | |  |  |  |  | **Activated carbon** | |  |  |
|  | (Control) | FIG  (2-.7MM) | FIG  (<2MM) | MAH  (2-.7MM) | MAH  (<2MM) | SHEA  (2-.7MM) | SHEA  (<2MM) | FIG  (2-.7MM) | FIG  (<2MM) | MAH  (2-.7MM) | MAH  (<2MM) | SHEA  (2-.7MM) | SHEA  (<2MM) |
| TOT – P | 40.22 | 4.81 | 5.36 | 5.26 | 6.13 | 5.41 | 6.07 | 11.12 | 13.67 | 11.31 | 14.31 | 11.74 | 14.04 |
| COD | 500 | 50 | 80 | 80.00 | 90.00 | 20.00 | 30.00 | 130.00 | 140.00 | 60.00 | 70.00 | 40.00 | 60.00 |
| NO3-N | 54.5 | 7 | 10.5 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 | 7 |
| pH | 89.49 | 6.34 | 6.58 | 6.37 | 6.29 | 6.4 | 6.7 | 6.77 | 7.11 | 7.31 | 7.51 | 7.49 | 7.51 |
| PO4-P | 123.27 | 14.74 | 16.43 | 16.12 | 18.79 | 16.58 | 18.6 | 34.08 | 41.90 | 34.67 | 43.80 | 35.98 | 43.03 |
| TOT-N | 35 | 7.00 | 7.00 | 7.00 | 7.00 | 7.00 | 7.0 | 7.00 | 7.00 | 7.00 | 7.00 | 7.00 | 7.00 |
| BOD | 270 | 50.00 | 60.00 | 100.00 | 110.00 | 80.00 | 80.0 | 70.00 | 80.00 | 70.00 | 70.00 | 90.00 | 90.00 |
| EC | 6092 | 1822 | 3490 | 2115 | 3001 | 1925 | 2431 | 1162 | 1483 | 1017 | 1078 | 1126 | 1163 |
| NH4-N | 1.8 | 1.12 | 0.96 | 0.88 | 1.04 | 0.96 | 0.16 | 0.20 | 0.12 | 0.20 | 0.20 | 0.16 | 0.52 |

Table 4.22 shows the measured level of pollutants at the end of the second week. All results are reported in MGL-1 except for pH which has no unit and EC which is reported in µS/cm.

## Table 4.23 Summary of Measured ParametersFor All Columns In Week 3

|  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Parameter** | **Initial**  **Reading** |  |  | **Biochar** | |  |  |  |  | **Activated carbon** | |  |  |
|  | (Control) | FIG  (2-.7MM) | FIG  (<2MM) | MAH  (2-.7MM) | MAH  (<2MM) | SHEA  (2-.7MM) | SHEA  (<2MM) | FIG  (2-.7MM) | FIG  (<2MM) | MAH  (2-.7MM) | MAH  (<2MM) | SHEA  (2-.7MM) | SHEA  (<2MM) |
| TOT – P | 37.11 | 2.64 | 4.77 | 3.85 | 6.59 | 3.64 | 6.82 | 8.86 | 10.59 | 6.59 | 9.82 | 7.15 | 8.32 |
| COD | 240 | 60 | 90 | 70 | 90 | 30 | 60 | 130 | 140 | 60 | 80 | 70 | 80 |
| NO3-N | 28 | 7 | 0 | 0 | 0 | 0 | 7 | 7 | 7 | 10.5 | 7 | 0.5 | 10.5 |
| pH | 7.93 | 7.31 | 7.46 | 7.23 | 7.61 | 7.41 | 7.32 | 7.82 | 7.91 | 7.66 | 7.57 | 7.63 | 8.14 |
| PO4-P | 113.74 | 8.09 | 14.62 | 11.86 | 20.20 | 11.16 | 20.90 | 27.16 | 32.46 | 20.21 | 30.10 | 21.91 | 23.50 |
| TOT-N | 21 | 7.00 | 7.00 | 7.00 | 7.00 | 7.00 | 7.00 | 7.00 | 7.00 | 7.00 | 7.00 | 7.00 | 7.00 |
| BOD | 350 | 80 | 80 | 110 | 110 | 110 | 140 | 120 | 120 | 100 | 110 | 120 | 130 |
| EC | 4412 | 2314 | 2518 | 1424 | 2497 | 1104 | 1588 | 818 | 1036 | 712 | 773 | 772 | 836 |
| NH4-N | 1.2 | 1.12 | 1.20 | 1.04 | 0.88 | 0.96 | 0.20 | 0.44 | 0.24 | 0.28 | 0.20 | 0.36 | 0.64 |

Table 4.23 shows the measured level of pollutants at the end of the third week. All results are reported in MGL-1 except for pH which has no unit and EC which is reported in µS/cm.

## Discussion of Results

* 1. **Greywater Influent Characteristics**

The greywater was found to be highly polluted than reported in similar studies. This was probably due to the non-regulation of products used in laundry and toilets such as phosphorus containing soap and detergents as the value of Total Phosphorus in the greywater source was 90mg/l which was a very high value when compared to other greywater reports (3.6mg/l Berger,(2012); 4 – 14mg/l Eriksson *et al.,* (2002). Organic matter load in the water was also high compared to other literatures. BOD and COD values were 450mg/l and 600mg/l respectively as compared to Friedler,(2004) with 377mg/l and 679mg/l. Friedler, (2004) also reported an electrical conductivity value of 1040µS/cm which was low compared to 7231μS/cm obtained in this study.

## Treatment Efficiency

* + 1. **Organic Matter**

There were no observed significant difference between filter media of different particle sizes. Activated carbon significantly reduced organic matter than biochar among columns with observed significant difference. Between the three parent material, the fig column had significant reduction in BOD in all the three weeks. It was observed that BOD reduction was steady in the first week, it increased in thesecond week and then decreased again in the third week .

High efficiencies were observed for both Activated Carbon (85 – 95%) and biochar (76 – 90%). Similar results were observed for BOD removal with efficiency range of 65 – 80%

for Activated Carbon and 68 – 75% for Biochar. Similar results were reported by Berger, (2012)

## Electrical Conductivity

There were no significant difference in electrical conductivity of the filters with different particle sizes. Remarkable significant differences was observed between biocharand activated carbon. Biochar showed higher efficiencies (85 – 90%) than Activated Carbon (65 – 81%). As reported by Amonette&Josheph, (2009), lower observed efficiencies of Activated Carbon may be attributed to acid used for the activation which might altered the mineral composition and functional chemical groups of the surface. This in turn might have altered the charge and surface activity of the surface hence its lower efficiency. The results in the activated carbon kept increasing from the first to the second week and then slightly reduced in the third week.

The fig column performed better than the shea tree and the Mahoganycolumn for both biochar and activated carbon in the first week but shea-butter column performed better in the second and third week than the other two parent materials.

## NH4-N Adsorption

There were no observed significant differences between the compared particle sizes. Comparing the activation status however, biochar showed higher efficiency (68 – 89%) than Activated Carbon which showed very low efficiency (36 – 89%). The shea tree showed the best remediation performance among the compared parent materials. Ammonium Nitrogen values in the first week was lower compared to the second and third

week. This formation of biofilm at the surface of the adsorbent might be the possible cause of this decrease as suggested by Moore *et al,* (2001).

## NO3-N Adsorption

The highest efficiency recorded in this study was for the removal of Nitrates where Activated Carbon had an efficiency range from 93 – 100% while biochar showed efficiency of 91 – 99%. Filters from the shea tree performed excellently for both Activated Carbon and biochar than the two other parent materials used.

The reduction rate of Nitrate as observed from the control sample indicates the instability of Nitrate which might have aided in the recorded high removal efficiency. Similarly, the performance of the shea tree column shows that adsorption is also another factor behind the reduction of Nitrate.

There was not observed significant differences between the compared particle sizes in the reduction of Nitrate from the greywater.

## pH Stabilization.

The initial pH of the greywater was qui*et al*kaline and remained so as shown the result from the control column. Filters from the biochar column slowly stabilized the pH during the three weeks duration while the activated Carbon filters rapidly acidified the greywater in the first week before stabilizing it.Reaction between the greywater and the activated carbon surface might be a possible cause for the rapid reduction in alkalinity of the

greywater in the first week as observed from the activated Carbon filters. This result however differs from that reported by Berger, (2012)

## Total Nitrogen Adsorption

Results obtained from adsorption of Total Nitrogen was steady throughout the experiment. Although the pollutants have been brought to an average stable value of 7.5mg/l, there were no observed significant difference between different particle sizes, parent material and even between biochar and activated carbon.

## Total Phosphorus Adsorption

Treatment efficiency of Total Phosphorus remained high throughout the experiment with activated Carbon having and efficiency range from 92 – 99% while biochar recorded 88 – 92%. The high efficiency recorded in the first week despite high control value suggest that adsorption might be the primary factor responsible for the reduction of Total Phosphorus in the greywater.

## PO4-P Adsorption

Although the initial phosphate value of the greywater was high, reduction efficiency remained high throughout the experiment. Removal efficiency of phosphate followed a similar pattern with that of Total Phosphorus which is inline with a similar study reported by PPRC, (2014).

## Summary of Discussion of Results

* + 1. **Effect of Particle Size**

Apart from the two instances in NH4N and Total-P, differences between the 2 – 4.7mm and

<2mm did not show significant difference for both Biochar and Activated Carbon. Although small differences were recorded, they were not qualified as statistically significant. The larger particle sizes (2 – 4.7mm) however performed better than the smaller particle sizes (<2mm) for both biochar and Activated Carbon. This can be attributed to the particle size distribution of the filter media. The larger sized filter (2- 4.7mm) had higher porosity hence more residence time, higher hydraulic conductivity and therefore more adsorption as reported by Dalameh, (2013).

## Effect of Parent Material

Among the three parent materials compared (Fig, Mahogany and Shea tree), Biochar and Activated Carbon from the shea tree recorded best performance in remediating COD, EC, NH4-N, PO4-P and TOT-P. Neutral performance was recorded in remediation of NO3-N and TOT-N. Filters from the Mahogany tree recorded best for pH stabilization for both Biochar and Activated Carbon. The Fig tree has the best performance in remediation of BOD.

These differences can be attributed to differences in the lignin content of the trees as suggested by Kearns*et al*., (2014).

## Effect of Activation

The activated carbon was observed to have better performance in remediation of COD, NO3-N, PO4-P, TOT-P, pH and BOD. Biochar however showed better performance in

remediating EC, NH4-N and Tot-N. Of the observed differences, only differences in COD, EC, PO4-N and NH4-N were observed to be statistically significant.

Differences in performance between the Biochar and the Activated Carbon may be attributed to the mineral composition and the functional chemical groups of the surface which determines charge and activity at the surface. This in turn affects the rate and type of pollutants to be adsorbed. The activation of the Activated Carbon with H2SO4 acid might have altered the surface composition (Amonette andJosheph, 2009)

## Effect of Time

Although better remediation was observed in the third week, the first week had the best remediation performance in terms of efficiency as the pollution load was more in the first week and significantly reduced in the two final weeks except for BOD which kept increasing. The first week recorded better performance in remediating NH4-N, NO3-N, PO4, TOT-N, TOT-P and pH. BOD was best remediated in the second week while Electrical Conductivity was best remediated in the third week.

This pattern agrees with the assumption of Trulear and Characklis,(1982), who suggested that adsorption is a prerequisite for microbial attachment where the medium host the microbes and the adsorbed nutrient provides the feed.

It also agrees with the Langmuir monolayer assumption as to adsorption.

## General Results Performance

Results from the experiment showed that all the filter column had significant performance in remediating the pollutants from the contaminated water. It also showed that there were variations in their treatment efficiency levels as time, parent material and activation status played a significant role in their performance. Phosphate Phosphorus, Ammonium Nitrogen and Total Phosphorus were better absorbed at the first week while most of the other parameters; COD, Electrical Conductivity, Nitrate-Nitrogen, BOD, Total-Nitrogen and pH showed acceptable concentration after the third week. The filter columns had remediated all the polluted water from all the tested parameters to levels that meets the effluent discharge standard in many countries and is even suitable for other purposes like unrestricted irrigation, reclaimed domestic use as seen in table 4.25

## TABLE 4.24 Treatment EffeciencyOf Filters

|  |  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **PARAMETER** |  |  | **ACTIVATED CARBON** | | |  |  |  | **BIOCHAR** | |  |  |
|  | FIG  (2-4.7) | FIG (<2) | MAHOG ANY  (2-4.7) | MAHOG ANY (<2) | SHEA 2-4.7 | SHEA  <2 | FIG  (2-4.7) | FIG (<2) | MAHOG ANY  (2-4.7) | MAHOG ANY (<2) | SHEA 2-4.7 | SHEA  <2 |
| **EC** | 81 | 65 | 80 | 65 | 85 | 78 | 88 | 85 | 90 | 89 | 89 | 88 |
| **TOT-P** | 97 | 94 | 96 | 92 | 96 | 92 | 90 | 88 | 93 | 89 | 92 | 91 |
| **NO3-N** | 93 | 100 | 100 | 100 | 100 | 94 | 94 | 94 | 91 | 94 | 99 | 91 |
| **BOD5** | 80 | 80 | 73 | 73 | 73 | 65 | 70 | 70 | 75 | 73 | 70 | 68 |
| **NH4-N** | 41 | 36 | 45 | 53 | 49 | 89 | 76 | 87 | 85 | 89 | 81 | 66 |
| **TOT-N** | 80 | 80 | 80 | 80 | 80 | 80 | 80 | 80 | 80 | 80 | 80 | 80 |
| **COD** | 90 | 85 | 88 | 85 | 95 | 90 | 78 | 76 | 90 | 86 | 88 | 86 |
| **PO4-P** | 97 | 95 | 96 | 93 | 96 | 92 | 90 | 88 | 93 | 89 | 92 | 91 |
| **pH** | - | - | - | - | - | - | - | - | - | - | - | - |

**TABLE 4.25 INTERNATIONAL WASTEWATER STANDARDS AND REUSE.**

|  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- |
| PARAMETER | INTERNATIONAL WASTE WATER RE-USE STANDARDS | | | |  |  |
|  | **FAO** | **EU** | **JORDAN** | **EGYPT** | **ITALY** | **CHINA** |
| EC (US/CM) | **3000** |  | **1500** |  |  |  |
| NO3-N (MG/L) | **30** |  |  |  |  |  |
| PH | **6.5-8.4** |  | **6 – 9** |  | **6 – 9.5** | **6 - 9** |
| NH4-N (MG/L) | **5** |  |  |  |  | **5** |
| BOD (MG/L) | **100** |  |  |  |  |  |
| TOT-N (MG/L) | **45** | **10 – 15** |  |  | **15** | **15** |
| PO4-P (MG/L) | **2** | **1 – 2** |  |  |  |  |
| TOT-P (MG/L) | **-** |  |  |  | **2** | **0.5** |
| COD (MG/L) | **150** | **125** | **100** | **40** | **100** |  |
| RE-USE | Unrestricted Irrigation | Discharge from urban wastewater treatment plants | Reclaimed Domestic reuse | Unrestricted Irrigation | Irrigation, urban Reuse | Impoundments And lakes |
| SOURCE | Ayers *et al* (1985) | Council directive (1991) | Dalahmeh*et al*.(2011) | Smith &Bani- Melhem (2012) | Chaillou*et al*.(2009) | Ernst *et al*. (2006) |

## CHAPTER FIVE

**SUMMARY, CONCLUSION AND RECOMMENDATION**

## Summary

The potential of biochar to remediate greywater was investigated. Greywater from Suleiman Hostel in Ahmadu Bello University was obtained and subjected to biocharfilters made from three different parent trees; *Vitellariaparadoxa* (Shea tree), *Khayasenegalenses* (Mahogany), *Ficussycomous*(Fig)for a period of 21 days. The biochar was prepared by slow pyrolysis of the trees and was used as filters in the experiment. The filters were classified into different sizes (range from 2mm to 4.7mm and another range of less than 2mm) to ascertain the effect of particle size on the efficiency of treatment. Same biochar of the same particle size were also activated using 20%H2SO4solution to produce activated carbon which was also utilized as filter media. Columns made from PVC pipes were designed,constructed and used as filter columns during the experiment. 2 liters of greywater (composite sample) was loaded in all the 13 columns (12 with filter media and one as control) at the beginning of the experiment and daily composite samples were also loaded daily to replace the quantity taken for laboratory analysis of basic greywater pollutants (EC, NH4-N, TOT-N, PO4-P, pH, NO3-N, TOT-P, BOD and COD). The

composite samples were collected at 8:00 AM and 4:00PM daily.

The results from the laboratory analysis were subjected to statistical analysis using Microsoft Excel 2013 and Graph Pad Instat version 3. Results from both biochar and activated carbon were compared and biochar shows better performance in remediating COD, BOD, Tot-Phosphorus, PO4-P and even in stabilizing the pH of the greywater. Activated carbon only performed better in remediating NH4-N and Electrical conductivity.

Neutral performance were observed from NO3-N and TOT-N adsorption. The performance efficiencies of both biochar and activated carbon in remediating greywater in this study was found to be similar with results from similar studies made by Berger, 2012.

## CONCLUSION

* + 1. In this study, a column adsorption of physic-chemical parameters of greywater was experimentally studied using biochar and activated carbon made from three different parent materials and their remediation potential was determined. The effect of particle sizes, activation status, parent materials and time was considered during the remediation process.
    2. Treatment efficiencies of both biochar and activated carbon was found to be high when compared to the results from the control column. Biochar however showed better performance in stabilizing pH, adsorption of PO4-P, TOT-P, BOD and COD. Neutral performance was observed in treating Total Nitrogen and NO3-N. Activated carbon performed better in reducing electrical conductivity and NH4-N.
    3. Production and handling of biochar was more cheaper and easier compared to the high cost of producing Activated Carbon which requires skilled knowledge handling and with the results obtained from this study, biochar can function as an activated carbon hence its better replacement.
    4. It was observed that the rate of adsorption between different particle sizes is heavily dependent on the porosity and distribution among the particle sizes than between the differences in the sizes itself.
    5. The filters made from the shea tree had the best performance followed by the Mahogany then the Fig. This suggest that Adsorption is possibly function of the woodiness or lignin content of the trees
    6. Adsorption in terms of efficiency was best observed in the first week and deteriorated in the second and third week for most of the filters. This suggest the formation of biofilm and possible compliance to the Langmuir‘s Isotherm model.

## RECOMMENDATION

* + 1. It is recommended that Biochar should be tried as an alternative to Activated Carbon in most treatment systems.
    2. Further studies should be conducted to observe the performance of Biochar under different Hydraulic Loading conditions.
    3. It is recommended that other tropical trees with higher lignin content should be investigated, charred and tried as filter media.
    4. It is also recommended that the used biochar to be tried for soil amendment potential.

## REFERENCES

*Ahmaruzzaman, M.D., (2008). Adsorption of phenolic compounds on low-cost adsorbents: a review, Adv. Colloid Interface Sci. 143 (1–2) 48–67.*

Agoyi, O.R., (2015). Performance of Cow bone char in Fix-bed column for the treatment of Industrial wastewater. Department of Water Resources and Environmental Engineering. Ahmadu Bello University. Pp 106-107.

Allen, L., Christian-Smith J., andPalaniappan M., (2010). Overview of Greywater Reuse: ThePotential of Greywater Systems to Aid Sustainable Water Management. Pacific Institute, Preservation Park, Oakland, California 94612, p. 40.

Amonette, J.E. andJosheph, S. (2009). Characteristics of Biochar: Microchemical Properties. *Biochar for environmental management science and technology*. London: Earthscan. ISBN 978-1-84407-658-1.

*Antal*, M.J., and *Gronli*, M., (*2003*). The Art, Science, and Technology of Charcoal Production. Industrial Engineering Chemistry Research, 42, 1619-1640.

Ayers, R.S., and Westcot D. W.,(1985). Water Quality for Agriculture. Food and Agricultural Organization of the United Nations, Rome

Aygun,A.,Yenisoy-Karakas,S.,and Duman,I.,(2003).Productionofgranular activated carbon fromfruitstonesandnutshellsandevaluationoftheir physical,chemical andadsorptionproperties. Journal of Soil biology and Chemistry Vol. 58,Pp140– 146.

Baldock, J. A. and Smernik, R. J. (2002). ‗Chemical composition and bioavailability of thermally altered *Pinusresinosa*(Red pine) wood‘, *Organic Geochemistry*, vol 33, pp1093–1109

*Barker, A.V., and English. J.E.,(2011).* [*"Recycling Gray Water for Home Gardens"*](http://extension.umass.edu/landscape/fact-sheets/recycling-gray-water-home-gardens)*.*

*University of Massachusetts. Archived from the original on September 1, 2012.*

*Berger, C., (2012). Biochar and Activated Carbon filters for greywater treatment. Comparison of organic matter and nutrient removal. Swedish University of Agricultural Sciences. ISSN1654-9392. Pp 5-16*

Bernd, M., Steffen W., Karsten A., and Manfred L., (2013).Potential dual use of biochar for wastewater treatment and soilamelioration Geophysical Research AbstractsVol. 15,

Pp 2214.

Bhatnagar, A., and Sillanpaa, M., (2010). Utilization of Agro-Industrial and Municipal Waste Materials as Potential Adsorbents for Water Treatment: A Review. *Chemical Engineering Journal*, 157, 277-296.

<http://dx.doi.org/10.1016/j.cej.2010.01.007>

Brinkmann, K., Blaschke, L., andPolle, A., (2002). Comparison of different methods for lignin determination as a basis for calibration of near-infrared reflectance spectroscopyand implications of lignoproteins. *J. Chem. Ecol.28*, 2483–2501.

Bulut, E., Ozacar M., and Sengil I.A.,(2008). Adsorption of malachite green onto bentonite: equilibrium and kinetic studies and process design, Micropor. Mesopor.Mater. 115 234–246.

Burnat, J.M.Y. and Mahmoud, N., (2005). Evaluation of On-Site Gray Wastewater TreatmentPlants Performance in Bilien and Biet-Diko Villages/Palestine, Environment ProtectionCommittee (EPC).

Chaillou, K., Gérente, C., Andrès, Y., and Wolbert, D. (2009). Bathroom greywater characterization and potential treatments for reuse. Water Air Soil Pollution. 215, 31-42.

Chan, K.Y., andXu, Z.,(2009). Biochar: Nutrient properties and their enrichment. Biochar for environmental management: Science and technology.pp.67-84

Clay, S.A., and Malo D.D., (2012). The Influence of Biochar Production on Herbicide Sorption Characteristics,Herbicides - Properties, Synthesis and Control of Weeds, ISBN: 978-953-307-803-8, InTech, Available from: <http://www.intechopen.com/books/herbicides-properties-synthesisand-control-of-> weeds/the-influence-of-biochar-production-on-herbicide-sorption-characteristics

Corcoran, E., Nellemann, C., Baker, E., Bos, R., Osborn, D. andSavelli, H. (2010). *Sick Water? The central role of wastewter management in sustainable development. A Rapid Responce Assessment*. As, Norway: United Nations Environment Programme, UN-HAPITAT, GRID-Arendal.

Council Directive (CD).,(1991).Concerning urban waste water teatment. Published at[:http://eur](http://eur-ex.europa.eu/LexUriServ/LexUriServ.do?uri=OEN)-[ex.europa.eu/LexUriServ/LexUriServ.do?uri=OEN](http://eur-ex.europa.eu/LexUriServ/LexUriServ.do?uri=OEN) (30.1.2016)

Dabrowski,A.,(2003). Adsorption—from theory to practice, Adv. Colloid Interface Sci.

93 135–224.

Dalahmeh, S.S., (2013). Bark and Charcoal filters for greywater treatment. Department of Energy and technology, Swedish University of Agricultural Sciences, Pp11-23

Dalahmeh, S.S., Hylander, L.D., Vinneras, B., Pell, M., Oborn, I., and Jonsson, H., (2011).

Potential of organic filter materials for treating greywater to achieve irrigation quality: a review. Water Science & Technology, 63,1832–1840.

Dalahmeh, S.S., Pell, M., Vinneras, B., Hylander, L.D., Oborn, and I., Jonsson, H., (2012).

Efficiency of bark, activated charcoal, foam and sand filters in reducing pollutants from greywater. Water, Air, & Soil Pollution, 1–15.

Davin, L.B.; Patten, A.M., Jourdes, M., and Lewis, N.G. (2008).Lignins: A twenty-first century challenge. In *Biomass Recaltritrance. Deconstructing the Plant Cell Wall forBioenergy*; Himmel, M.E., Ed.; Blackwell Publishing Ltd.: Oxford, UK, pp.

213–305.

Demirbas, A. (2004). ‗Bioenergy, global warming, and environmental impacts‘, *Energy Sources*, vol 26, pp225–236

Downie, A., Krosky, A. and Munroe, P. (2009). Physical Properties of Biochar. In: *Biochar for environmental management science and technology*. London: Earthscan. ISBN 978-1-84407-658-1.

Eriksson, E., Auffarth, K., Henze, M., andLedin, A.,(2002).Characteristics of grey wastewater. Urban Water, 4 (1), 85-104.

Ernst, M., Sperlich, A., Zheng, X., Gan, Y., Hu, J., Zhao, X., Wang, J., Jekel, M.,(2007). An Integrated wastewater treatment and reuse concept for the Olympic Park 2008, Beijing.Desalination, 202(1-3), 293–301.

Faruqui, N., and Al-Jayyousi, O., (2002). Greywater reuse in urban agriculture for poverty alleviation - A case study in Jordan. Water International, 27(3): 387-394.

Foereid, B., (2015).Biochar in Nutrient Recycling:The Effect and Its Use in Wastewater Treatment Norwegian Institute for Agricultural and Environmental Research.

Foo, K.Y., Hameed B.H., (2010). Insights into the modelling of Adsorption isotherm systems. Journal of Chemical Engineering 156 2-10

Friedler, E. (2008). The water saving potential and the socio-economic feasibility of greywater reuse within the urban sector - Israel as a case study. International Journal ofEnvironmental Studies, 65: 1, 57 - 69.

Friedler, E., (2004). Quality of individual domestic greywater streams and its implication for onsite treatment and reuse possibilities. Environmental Technology, 25(9): 997-

1008.

Friedler, E. and Galil N. I., (2003). On-site greywater reuse in multi-storey buildings: Sustainable solution for water saving. *Efficient 2003 - 2nd Int. Conf. on Efficient Use and Management of Urban Water Supply*. Tenerife, Canary Islands Spain, April 2003.

Geoffery, L., ( 2008). "Ancient skills 'could reverse global warming'" The Independent.

Retrieved 1 October 2011.

Ghezzehei, T. A.,Sarkhot, D.V., and Berhe A.A.,(2014).Biochar can be used to capture essential nutrients from dairy wastewater and improve soil physico-chemical properties. Solid Earth Discuss.

Gross, A., Shmueli, O., Ronen, Z. and Raveh, E., (2006). Recycled Vertical Flow ConstructedWetland (RVFCW) - a novel method of recycling greywater for landscape irrigation in smallcommunities and households. Chemosphere..

Guerra A., Filpponen I., Lucia L.A., Saquing C., Baumberger S., and Argyropoulos D.S., (2006).Toward a better Understanding of the Ligning Isolation Process from Wood. J. Agric. Food Chem. 2006, 54, 5939-5947

Harris, P. (1999). ‗On charcoal‘, *Interdisciplinary Science Reviews*, Vol 24, pp301–306 Harkin, J.M.,(1969) Ligning and its uses. U.S Forest Service Research Note FPL-0206

Hillel, D., (1982) Introduction to Soil Physics. *Department of plant and soil science.*

*University of Masachusset. Pp 20-44*

Huggins, T. M.,Haeger A.,BiffingerJ.C., and Ren, Z J., (2016)"Granular biochar compared with activated carbon for wastewater treatment and resource recovery". *U.S. Navy Research.* Paper 99.

Igboro, S.B., Alfa M.I., Ismail A., Dahunsi S.O., and Komami M., (2016) Evaluation of the efficiency of PakiaBiglobosa (locust bean) seed, leaf and bark extracts for the treatment of greywater. Nigerian Journal of Scientific Research 15(1) Pp 75-80

Igboro, S.B., Ezenwe U.C., Nwude M.O., and Fumen G.A., (2015) Reuse of greywater for theproduction of spinach (Amaranthusspp) in Semi-Arid Region of Nigeria. *NigerianJournal of Hydrological Sciences*. Vol 3: 1-8. Pp 2-7

Kearns, J., (2012) Sustainable Decentralized Water Treatment for Rural and Developing Communities Using Locally Generated Biochar Adsorbents.

Aqsolutions Pp 1-5

Kearns J., Wellborn L.S., Summers R.S.,andKnappeD.R.. (2014) 2,4-D adsorption to biochars: Effect of preparation conditions on equilibrium adsorption capacity and comparison with commercial activated carbon literature data. *Water Research*, Vol. 62, pp. 20-28, 2014.

Khan, M.A., Kim, S.W., Khan-Rao A.R., Abou-Shanab R.A.,Bhatnagar A., Song H.,Jeon

H. B.,(2010) Adsorption studies of Dichloromethane on some commercially available GACs: Effect of kinetics, thermodynamics and competitive ions. *Journal ofHazardous Materials.* Vol. 178. 963-972

Krull, E.S., BaldockJ.A., SkjemstadJ.O., andSmernik R.J.,(2009). Characteristics of biochar: Organochemical properties. pp. 53-65. In J. Lehmann and S. Joseph (eds.) Biochar for environmentalmanagement: *Science and technology. Earthscan, London.*

Kundu S., and Gupta A.K.,(2006) Arsenic adsorption onto iron oxide-coated cement (IOCC): regression analysis of equilibrium data with several isotherm models and their optimization, *Chem. Eng. J*. 122 93–106.

Laird, D.A., (2008) ‗The charcoal vision:A win–win–win scenario for simultaneously producing bioenergy, permanently sequestering carbon, while improving soil and waterquality‘, *Agronomy Journal*, vol 100, pp178–181

*Langmuir*, I., (*1916*) The constitution and fundamental properties of solids and liquids.

*Journal of American Chemistry Society*, 38(11), 2221-2221.

Lehman J., and Joseph S., (2009). Biochar for Environmental Management: An Introduction. *Frontiers in ecology and the Environment 16(2). Pp 1-7.*

Li, F., Wichmann, K. and Otterpohl, R. (2009) Review of the technological approaches for grey water treatment and reuses. Science of the Total Environment 407, 3439–3449.

Li, M.Y., Zhang, J., Lu, P., Xu, J.L., and Li, S.P., (2009). Evaluation of Biological Characteristics of Bacteria Contributing to Biofilm Formation. *Pedosphere*19(5), 554-561.

Liang, B., Lehmann, J., Solomon, D., Kinyangi, J., Grossman, J., O‘Neill, B., Skjemstad, J. O., Thies, J., Luizao, F. J., Petersen, J.,and Neves, E. G.(2006). Black carbon increases cation exchange capacity in soils, *Soil Sci. Soc. Am. J*., 70, 1719– 1730, 2006.

Limousin G., Gaudet J.P., Charlet L., Szenknect S., Barthes V., and Krimissa M.,(2007) SorptionIsotherms: a review on physical bases, modeling and measurement, *Appl*. *Geochem.* 22 249–275.

Ludwig, A., (2012). Create an oasis of greywater: Choosing, using and building greywater systems. Oasis Design Santa Barbara C.A Pp 1-2

Malek, A., and Farooq S.,(1996) Comparison of isotherm models for hydrocarbon adsorptionon activated carbon, *AIChE* J. 42 (11) 3191–3201.

Martin, C., (2005). Ecological Sanitation greywater demonstration project at Hui Sing garden,Kuching, Sarawak, Malaysia, Urban Environmental Management System (UEMS)ProjectNatural Resources and Environment Board (NREB), Sarawak.

McKay, D.M., and Roberts P.V.,(1982). The dependence of char and carbon yield on lignocellulosicprecursor composition. Carbon. 20:87.

Mclaughlin H., Anderson P.S., Shields F.E., and Reed T.B., (2009): All Biochars are not created equal and how to tell them apart. AlternaBiocarbon Inc. Pp 1-7

Moore, R., Quarmby, J. and Stephenson, T., (2001). The effects of media size on the performance of biological aerated filters. *Water Research* 35(10), 2514-2522.

Morel A., andDiener S., (2006). Greywater management in low and middle-income countries, review of different treatment systems for households or neighbourhoods. Dübendorf:*Swiss Federal Institute of Aquatic Science and Technology (Eawag) National Institue for Environmental Health Sciences (NIEHS) 2013*

Mungai G.G. (2008). Impacts of long-term greywater disposal on soil properties and reuse inurban agriculture in an informal settlement – a case study of Waruku, Nairobi.

*UNESCO-IHE Institute for Water Education, Eawag*.

Murphy K. O'H., (2006). A Scoping Study to Evaluate the Fitness-for-use of Greywater in Urban and Peri-Urban Agriculture*.* Pretoria: Water Research Commission Report. 1479/1/06. 1-77005-471-5.

Oron G., Adel M., Agmon, V., Friedler E., Halperin R., Leshem E., and Weinberg D., (2014)**. Greywater use in Israel and worldwide: standards and prospects.** *Water Resources*, **58**: 92-101.

Orwa, C.; Mutua, A.; Kindt, R.; Jamnadass, R.; andAnthony, S., (2009). Agroforestree Database: a tree reference and selection guide version 4.0. *World Agroforestry Centre, Kenya*.

Padmesh T.V.N., Vijayaraghavan K., Sekaran G., Velan M., (2006) Application of two and three parameters isotherm models: Biosorption of acid red 88 onto *Azolla microphylla*. Bioremediation Journal, Vol. 10, No. 1-2, pp. 37-44.

Pérez-Marín A.B., Zapata V.M, Ortuno J.F., Aguilar M., Sáez J., andLlorens M., (2007) Removal of cadmium from aqueous solutions by adsorption onto orange waste, J. Hazard. Mater. B139 122–131.

Pollution Prevention Resource Centre (PPRC)., (2014), Emerging Best Management Practices in Stormwater:Biochar as filtermedia:<http://pprc.org/wp-> content/uploads/2014/08/Emerging-Stormwater-BMPs\_Biochar-as-Filtration- media\_2014.pdf.

Redwood, M., (2004). Wastewater use in urban agriculture, International Development ResearchCentre IDRC.

Schwarzenbach, R.P., Escher, B.I.; Fenner K., Hofstetter T.B., Annette Johnson, C.A., Von G., and Wehrli, B., (2006). The Challenge of Micropollutants inAquatic Systems. Science, Vol. 313, p. 1072.

Schnizter, M.I., Monreal C., Jandl G., Leinweber P., andFransham. P.B.,(2007). The conversion of chickenmanure to biooil by fast pyrolysis: II. Analysis of Chicken

manure, biooils, and char by curie-pointpyrolysis-gas chromatography/mass spectrometry. J, Environ. Sci. Health B. 42:79-95.

Shrestha, R.R., Haberl, R., Laber, J., Manandhar, R. and Mader, J., (2001). Application of constructed wetlands for wastewater treatment in Nepal. Water Science and Technology,44(11-12): 381-386

Sluiter, A., Hames, B.,Ruiz, R.,Scarlata, C., Sluiter, J., and Templeton, D., (2008) Determination of structural carbohydrates and lignin in biomass: laboratory analytical procedure (LAP). Golden, CO: National Renewable Energy Laboratory.

Smith, E., and Bani-Melhem, K., (2012). Grey water characterization and treatment for reuse in an arid environment.Water Science & Technology, 66(1), 72-78.

Steinbiss, S., Gleixner, G. and Antonietti, M. (2009) Effect of Biochar on Soil Carbon Balance and Soil Microbial Activity. *Soil Biology & Biochemistry*, 41, 1301-1310. <http://dx.doi.org/10.1016/j.soilbio.2009.03.016>

Taghizadeh-Toosi, A., Clough, T.J., Sherlock, R.R. and Condron, L.M. (2012) A Wood Based Low Temperature Biochar Captures NH3-N Generated from Ruminant

Urine- N,Retaining Its Bioavailability. *Plant and Soil*, 353, 73-84.

Trulear, M.G.,andCharacklis, W.G., (1982). Dynamics of Biofilm Processes. *Journal of Water Pollution Control Federation* 54(9), 1288-1301.

United State Department of Statistics.,(2001). Jordan in Figures. In: Department of Statistics (Editor). Hashemite Kingdom ofJordan.

UN-Water Report (2006). *Water: A shared responsibility*. Paris. (The United Nations World Water Development. ISSN Report 2.

Vijayaraghavan K., Padmesh T.V., Palanivelu K., and Velan M.,(2006) Biosorption of Nickel(II) ions onto Sargassumwightii: application of two-parameter and three parameter isotherm models, J. Hazard. Mater. B133 304–308.

Wesseling, C., (1997) Agricultural Pesticide Use in Developing Countries: Health Effects and Research Needs.‖ International Journal of Health Services, 27.2 (1997): 273-308.

Winblad, U., and Simpson H, M., (2004). Ecological Sanitation – Revised and Enlarged Edition. Stockholm, Stockholm Environment Institute.Pp 22-23

World Health Organization Report (2009). Ten Facts About Water Scarcity.

Available from: [http://www.who.int/features/factfiles/water/en/index.html.](http://www.who.int/features/factfiles/water/en/index.html) [13 April 2014].

World Health Organization and UNICEF (2010), Progression Sanitation and Drinking Water, 2010 Update.

Zabaniotou, A., Stavropoulos G., and Skoulou V.,(2008). Activated carbon from olive kernels in a twostageprocess: Industrial improvement. Biores. Technol. 99:320- 326.

Zohary D., and Maria H., (2000) *Domestication of plants in the Old World*, third edition (Oxford:University Press), p. 165

APPENDIX A

## APPENDICES

**PICTURE OF CONSTRUCTED FILTER COLUMNS**

 

**Pictures of sealed Filter Columns before loading Crossection of filter columns on stand**



**Picture of unsealed columns during loading Crossection of filter columns on stand.**

APPENDIX B

United States Department of Energy Method of determining Lignin

0.3 g of dried extracted raw biomass was weighed in glass test tubes and 3 ml of 72% H2SO4 was added. The sample was kept at room temperature for 2 hours with carefully shaking at 30 min intervals to allow for complete hydrolysis. After the initial hydrolysis, 84 ml of distilled water was added. The second step of hydrolysis was made to occur in an autoclave for 1 h at 121 °C. The slurry was then cooled at room temperature. Hydrolyzates were filtered through vaccum using a filtering crucible. The acid insoluble lignin was determined by drying the residues at 105 oC and accounting for ash by incinerating the hydrolyzed samples at 575 oC in a muffle furnace. The acid soluble lignin fraction was determined by measuring the absorbance of the acid hydrolyzed samples at 320 nm. The lignin content was calculated as the summation of acid insoluble lignin and acid soluble lignin.